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- (54) Title: ADENO-ASSOCIATED VIRUS SEROTYPE 1 NUCLEIC ACID SEQUENCES, VECTORS AND HOST CELLS CONTAINING SAME
- (54) Titre: SEQUENCES D'ACIDE NUCLEIQUE DU SEROTYPE I DU VIRUS ASSOCIE AUX ADENOVIRUS, VECTEURS ET CELLULES HOTES CONTENANT CES DERNIERS

(57) Abstract

The nucleic acid sequences of adeno-associated virus (AAV) serotype 1 are provided, as are vectors and host cells containing these sequences and functional fragments thereof. Also provided are methods of delivering genes via AAV-1 derived vectors.

(57) Abrégé

L'invention concerne des séquences d'acide nucléique du sérotype 1du virus associé aux adénovirus (AAV) ainsi que des vecteurs et des cellules hôtes contenant ces séquences et des fragments fonctionnels de ces derniers. L'invention traite également de procédés d'administration de gènes via des vecteurs dérivés de l'AVV-1.

CORRECTED **VERSION***



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(54) Title: ADENO-ASSOCIATED VIRUS SEROTYPE 1 NUCLEIC ACID SEQUENCES, VECTORS AND HOST CELLS CONTAIN-ING SAME

(57) Abstract

The nucleic acid sequences of adeno-associated virus (AAV) serotype 1 are provided, as are vectors and host cells containing these sequences and functional fragments thereof. Also provided are methods of delivering genes via AAV-1 derived vectors.

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Description

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ADENO-ASSOCIATED VIRUS SEROTYPE I NUCLEIC ACID SEQUENCES, VECTORS AND HOST CELLS CONTAINING SAME

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This work was supported by the National Institutes of Health, grant no. P30 DK47757-06 and PO1 HD32649-04. The US government may have certain rights in this invention.

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Field of the Invention

This invention relates generally to viral vector, and more particularly, to recombinant viral vectors useful for gene delivery.

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Background of the Invention

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Adeno-associated viruses are small, single-stranded DNA viruses which require helper virus to facilitate efficient replication [K.I. Berns, Parvoviridae: the viruses and their replication, p. 1007-1041, in F.N. Fields et al., Fundamental virology, 3rd ed., vol. 2, (Lippencott-Raven Publishers, Philadelphia, PA) (1995)]. The 4.7 kb genome of AAV is characterized by two inverted terminal repeats (ITR) and two open reading frames which encode the Rep proteins and Cap proteins, respectively. The Rep reading frame encodes four proteins of molecular weight 78 kD, 68 kD, 52 kD and 40 kD. These proteins function mainly in regulating AAV replication and integration of the AAV into a host cell's chromosomes. The Cap reading frame encodes three structural proteins in molecular weight 85 kD (VP 1), 72 kD (VP2) and 61 kD (VP3) [Berns, cited above]. More than 80% of total proteins in AAV virion comprise VP3. The two ITRs are the only cis elements essential for AAV replication, packaging and integration. There are two conformations of AAV ITRs called "flip" and "flop". These differences in conformation originated from the replication model of adeno-associated virus which use the ITR to initiate and reinitiate the replication [R.O. Snyder et al., J. Virol., 67:6096-6104 (1993); K.I. Berns. Microbiological Reviews, 54:316-329 (1990)].

AAVs have been found in many animal species, including primates, canine, fowl and human [F.A. Murphy et al., "The Classification and Nomenclature of Viruses: Sixth Report of the International Committee on Taxonomy of Viruses",

Archives of Virology, (Springer-Verlag, Vienna) (1995)]. In addition to five known primate AAVs (AAV-1 to AAV-5), AAV-6, another serotype closely related to AAV-2 and AAV-1 has also been isolated [E. A. Rutledge et al., J. Virol., 72:309-319 (1998)]. Among all known AAV serotypes, AAV-2 is perhaps the most well-characterized serotype, because its infectious clone was the first made [R.J. Samulski et al., Proc. Natl, Acad. Sci. USA, 79:2077-2081 (1982)]. Subsequently, the full sequences for AAV-3A, AAV-3B, AAV-4 and AAV-6 have also been determined [Rutledge, cited above; J.A.Chiorini et al., J. Virol., 71:6823-6833 (1997); S. Muramatsu et al., Virol., 221:208-217 (1996)]. Generally, all AAVs share more than 80% homology in nucleotide sequence.

A number of unique properties make AAV a promising vector for human gene therapy [Muzyczka, Current Topics in Microbiology and Immunology, 158:97-129 (1992)]. Unlike other viral vectors, AAVs have not been shown to be associated with any known human disease and are generally not considered pathogenic. Wild type AAV is capable of integrating into host chromosomes in a site specific manner [R. M. Kotin et al., Proc, Natl. Acad. Sci. USA, 87:2211-2215 (1990)- R.J. Samulski, EMBO J., 10(12):3941-3950 (1991)]. Recombinant AAV vectors can integrate into tissue cultured cells in chromosome 19 if the rep proteins are supplied in trans [C. Balague et al., J. Virol., 71:3299-3306 (1997); R. T. Surosky et al., J. Virol., 71:7951-7959 (1997)]. The integrated genomes of AAV have been shown to allow long term gene expression in a number of tissues, including, muscle, liver, and brain [K. J. Fisher, Nature Med., 3(3):306-312 (1997); R. 0. Snyder et al., Nature Genetics, 16:270-276 (1997); X. Xiao et al., Experimental Neurology, 144:113-124 (1997); Xiao, J. Virol., 70(11):8098-8108 (1996)].

AAV-2 has been shown to be present in about 80-90% of the human population. Earlier studies showed that neutralizing antibodies for AAV-2 are prevalent [W. P. Parks et al., <u>J. Virol.</u>, <u>2</u>:716-722 (1970)]. The presence of such antibodies may significantly decrease the usefulness of AAV vectors based on AAV-2 despite its other merits. What are needed in the art are vectors characterized by the

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advantages of AAV-2, including those described above, without the disadvantages, including the presence of neutralizing antibodies.

Summary of the Invention

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In one aspect, the invention provides an isolated AAV-1 nucleic acid molecule which is selected from among SEQ ID NO: 1, the strand complementary to SEQ ID NO: 1, and cDNA and RNA sequences complementary to SEQ ID NO: 1 and its complementary strand.

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In another aspect, the present invention provides AAV ITR sequences, which include the 5' ITR sequences, nt 1 to 143 of SEQ ID NO: 1; the 3' ITR sequences, nt 4576 to 4718 of SEQ ID NO: 1, and fragments thereof.

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In yet another aspect, the present invention provides a recombinant vector comprising an AAV-1 ITR and a selected transgene. Preferably, the vector comprises both the 5' and 3' AAV-1 ITRs between which the selected transgene is located.

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In still another aspect, the invention provides a recombinant vector comprising an AAV-1 P5 promoter having the sequence of nt 236 to 299 of SEQ ID NO: 1 or a functional fragment thereof.

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In a further aspect, the present invention provides a nucleic acid molecule encoding an AAV-1 rep coding region and an AAV-1 cap coding region.

In still another aspect, the present invention provides a host cell transduced with a recombinant viral vector of the invention. The invention further provides a host cell

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In still a further aspect, the present invention provides a pharmaceutical composition comprising a carrier and a vector of the invention.

stably transduced with an AAV-1 P5 promoter of the invention.

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In yet another aspect, the present invention provides a method for AAV-mediated delivery of a transgene to a host involving the step of delivering to a selected
host a recombinant viral vector comprising a selected transgene under the control of
sequences which direct expression thereof and an adeno-associated virus 1 (AAV-1)
virion.

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In another aspect, the invention provides a method for in vitro production of a selected gene product using a vector of the invention.

Other aspects and advantages of the invention will be readily apparent to one of skill in the art from the detailed description of the invention.

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Brief Description of the Drawings

Figs. 1A-1C illustrate the alignment of nucleotides of AAV-1 [SEQ ID NO: 1], AAV-2 [SEQ ID NO: 18] and AAV-6 [SEQ ID NO: 19]. The alignment was done with MacVector 6.0. The full sequences of AAV-1 are shown in the top line. Nucleotides in AAV-2 and AAV-6 identical to AAV-1 are symbolized by "." and gaps by "-". Some of the conserved features among AAVs are marked in this figure. Note the 3' ITRs of AAV-1 and AAV-6 are shown in different orientations.

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Fig. 2 illustrates the predicted secondary structure of AAV-1 ITR. The nucleotides in AAV-2 and AAV-6 are shown in italic and bold respectively.

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Fig. 3A illustrates a hypothesis of how AAV-6 arose from the homologous recombination between AAV-1 and AAV-2. The major elements of AAV-1 are indicated in the graph. A region that is shared between AAV-1, AAV-2 and AAV-6 is shown in box with waved lines.

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Fig. 3B is a detailed illustration of a 71 bp homologous region among AAV-1, AAV-2 and AAV-6. Nucleotides that differ among these serotypes are indicated by arrows.

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Fig. 4A is a bar chart illustrating expression levels of human alpha 1 antitrypsin (a1AT) in serum following delivery of hAAT via recombinant AAV-1 and recombinant AAV-2 viruses.

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Fig. 4B is a bar chart illustrating expression levels of erythropoietin (epo) in serum following delivery of the epo gene via recombinant AAV-1 and recombinant AAV-2 viruses

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Fig. 5A is a bar chart illustrating expression levels of $\alpha 1AT$ in liver following delivery of $\alpha 1AT$ as described in Example 7.

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5 Fig. 5B is a bar chart demonstrating expression levels of epo in liver following 10 delivery of epo as described in Example 7. Fig. 5C is a bar chart demonstrating neutralizing antibodies (NAB) directed to AAV-1 following delivery of a1AT or epo to liver as described in Example 7. Fig. 5D is a bar chart demonstrating neutralizing antibodies (NAB) directed to 5 15 AAV-2 following delivery of a IAT or epo to liver as described in Example 7. Fig. 6A is a bar chart illustrating expression levels of a IAT in muscle following delivery of a lAT as described in Example 7. 20 Fig. 6B is a bar chart demonstrating expression levels of epo in muscle 10 following delivery of epo as described in Example 7. Fig. 6C is a bar chart demonstrating neutralizing antibodies (NAB) directed to AAV-1 following delivery of a1AT or epo to muscle as described in Example 7. 25 Fig. 6D is a bar chart demonstrating neutralizing antibodies (NAB) directed to AAV-2 following delivery of a1AT or epo to muscle as described in Example 7. 30 15 Detailed Description of the Invention The present invention provides novel nucleic acid sequences for an adenoassociated virus of serotype 1 (AAV-1). Also provided are fragments of these AAV-1 sequences. Among particularly desirable AAV-1 fragments are the inverted terminal 35 repeat sequences (ITRs), rep and cap. Each of these fragments may be readily 20 utilized, e.g., as a cassette, in a variety of vector systems and host cells. Such fragments may be used alone, in combination with other AAV-1 sequences or 40 fragments, or in combination with elements from other AAV or non-AAV viral sequences. In one particularly desirable embodiment, a cassette may contain the AAV-1 ITRs of the invention flanking a selected transgene. In another desirable embodiment, a cassette may contain the AAV-1 rep and/or cap proteins, e.g., for use 25

in producing recombinant (rAAV) virus.

Thus, the AAV-I sequences and fragments thereof are useful in production of rAAV, and are also useful as antisense delivery vectors, gene therapy vectors, or vaccine vectors. The invention further provides nucleic acid molecules, gene delivery

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vectors, and host cells which contain the AAV-1 sequences of the invention. Also provided a novel methods of gene delivery using AAV vectors.

As described herein, the vectors of the invention containing the AAV-1 capsid proteins of the invention are particularly well suited for use in applications in which the neutralizing antibodies diminish the effectiveness of other AAV serotype based vectors, as well as other viral vectors. The rAAV vectors of the invention are particularly advantageous in rAAV readministration and repeat gene therapy.

These and other embodiments and advantages of the invention are described in more detail below. As used throughout this specification and the claims, the term "comprising" is inclusive of other components, elements, integers, steps and the like.

I. AAV-1 NUCLEIC ACID AND PROTEIN SEQUENCES

The AAV-1 nucleic acid sequences of the invention include the DNA sequences of SEQ ID NO: 1 (Figs. 1A-1C), which consists of 4718 nucleotides. The AAV-1 nucleic acid sequences of the invention further encompass the strand which is complementary to SEQ ID NO: 1, as well as the RNA and cDNA sequences corresponding to SEQ ID NO: 1 and its complementary strand. Also included in the nucleic acid sequences of the invention are natural variants and engineered modifications of SEQ ID NO: 1 and its complementary strand. Such modifications include, for example, labels which are known in the art, methylation, and substitution of one or more of the naturally occurring nucleotides with an analog.

Further included in this invention are nucleic acid sequences which are greater than 85%, preferably at least about 90%, more preferably at least about 95%, and most preferably at least about 98 - 99% identical or homologous to SEQ ID NO.1. The term "percent sequence identity" or "identical" in the context of nucleic acid sequences refers to the residues in the two sequences which are the same when aligned for maximum correspondence. The length of sequence identity comparison may be over the full-length sequence, or a fragment at least about nine nucleotides, usually at least about 20 - 24 nucleotides, at least about 28 - 32 nucleotides, and preferably at least about 36 or more nucleotides. There are a number of different

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algorithms known in the art which can be used to measure nucleotide sequence identity. For instance, polynucleotide sequences can be compared using Fasta, a program in GCG Version 6.1. Fasta provides alignments and percent sequence identity of the regions of the best overlap between the query and search sequences (Pearson, 1990, herein incorporated by reference). For instance, percent sequence identity between nucleic acid sequences can be determined using Fasta with its default parameters (a word size of 6 and the NOPAM factor for the scoring matrix) as provided in GCG Version 6.1, herein incorporated by reference.

The term "substantial homology" or "substantial similarity," when referring to a nucleic acid or fragment thereof, indicates that, when optimally aligned with appropriate nucleotide insertions or deletions with another nucleic acid (or its complementary strand), there is nucleotide sequence identity in at least about 95 - 99% of the sequence.

Also included within the invention are fragments of SEQ ID NO: 1, its complementary strand, cDNA and RNA complementary thereto. Suitable fragments are at least 15 nucleotides in length, and encompass functional fragments which are of biological interest. Certain of these fragments may be identified by reference to Figs. 1A-1C. Examples of particularly desirable functional fragments include the AAV-1 inverted terminal repeat (ITR) sequences of the invention. In contrast to the 145 nt ITRs of AAV-2, AAV-3, and AAV-4, the AAV-1 ITRs have been found to consist of only 143 nucleotides, yet advantageously are characterized by the T-shaped hairpin structure which is believed to be responsible for the ability of the AAV-2 ITRs to direct site-specific integration. In addition, AAV-1 is unique among other AAV serotypes, in that the 5' and 3' ITRs are identical. The full-length 5' ITR sequences of AAV-1 are provided at nucleotides I-143 of SEQ ID NO: 1 (Fig. 1A) and the fulllength 3' ITR sequences of AAV-1 are provided at nt 4576-4718 of SEQ ID NO: 1 (Fig. 1C). One of skill in the art can readily utilize less than the full-length 5' and/or 3' ITR sequences for various purposes and may construct modified ITRs using conventional techniques, e.g., as described for AAV-2 ITRs in Samulski et al, Cell, 33:135-143 (1983).

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Another desirable functional fragment of the AAV-1 genome is the P5 promoter of AAV-1 which has sequences unique among AAV P5 promoters, while maintaining critical regulatory elements and functions. This promoter is located within nt 236 - 299 of SEQ ID NO: 1 (Fig. 1A). Other examples of functional fragments of interest include the sequences at the junction of the rep/cap, e.g., the sequences spanning nt 2306-2223, as well as larger fragments which encompass this junction which may comprise 50 nucleotides on either side of this junction. Still other examples of functional fragments include the sequences encoding the rep proteins. Rep 78 is located in the region of nt 334 - 2306 of SEQ ID NO: 1; Rep 68 is located in the region of nt 334-2272, and contains an intron spanning nt 1924-2220 of SEQ ID NO: 1. Rep 52 is located in the region of nt 1007 - 2304 of SEQ ID NO: 1; rep 40 is located in the region of nt 1007 - 2272, and contains an intron spanning nt 1924-2246 of SEQ ID NO: 1. Also of interest are the sequences encoding the capsid proteins, VP 1 [nt 2223-4431 of SEQ ID NO: 1], VP2 [nt 2634-4432 of SEQ ID NO: 1] and VP3 [nt 2829-4432 of SEQ ID NO: 1]. Other fragments of interest may include the AAV-1 P19 sequences, AAV-1 P40 sequences, the rep binding site, and the terminal resolute site (TRS).

The invention further provides the proteins and fragments thereof which are encoded by the AAV-1 nucleic acids of the invention. Particularly desirable proteins include the rep and cap proteins, which are encoded by the nucleotide sequences identified above. These proteins include rep 78 [SEQ ID NO:5], rep 68 [SEQ ID NO:7], rep 52 [SEQ ID NO:9], rep 40 [SEQ ID NO: 11], vpl [SEQ ID NO: 13], vp2 [SEQ ID NO: 15], and vp3 [SEQ IID NO: 17] and functional fragments thereof while the sequences of the rep and cap proteins have been found to be closely related to those of AAV-6, there are differences in the amino acid sequences (see Table 1 below), as well as differences in the recognition of these proteins by the immune system. However, one of skill in the art may readily select other suitable proteins or protein fragments of biological interest. Suitably, such fragments are at least 8 amino acids in length. However, fragments of other desired lengths may be readily utilized.

The sequences, proteins, and fragments of the invention may be produced by

chemical synthesis.

any suitable means, including recombinant production, chemical synthesis, or other synthetic means. Such production methods are within the knowledge of those of skill in the art and are not a limitation of the present invention.

Such fragments may be produced recombinantly or by other suitable means, e.g.,

II. VIRAL VECTORS

In another aspect, the present invention provides vectors which utilize the AAV-1 sequences of the invention, including fragments thereof, for delivery of a heterologous gene or other nucleic acid sequences to a target cell. Suitably, these heterologous sequences (i.e., a transgene) encode a protein or gene product which is capable of being expressed in the target cell. Such a transgene may be constructed in the form of a "minigene". Such a "minigene" includes selected heterologous gene sequences and the other regulatory elements necessary to transcribe the gene and express the gene product in a host cell. Thus, the gene sequences are operatively linked to regulatory components in a manner which permit their transcription. Such components include conventional regulatory elements necessary to drive expression of the transgene in a cell containing the viral vector. The minigene may also contain a selected promoter which is linked to the transgene and located, with other regulatory elements, within the selected viral sequences of the recombinant vector.

Selection of the promoter is a routine matter and is not a limitation of this invention. Useful promoters may be constitutive promoters or regulated (inducible) promoters, which will enable control of the timing and amount of the transgene to be expressed. For example, desirable promoters include the cytomegalovirus (CMV) immediate early promoter/enhancer [see, e.g., Boshart et al, Cell, 41:521-530 (1985)], the Rous sarcoma virus LTR promoter/enhancer, and the chicken cytoplasmic β-actin promoter [T. A. Kost et al, Nucl, Acids Res., 11(23):8287 (1983)]. Still other desirable promoters are the albumin promoter and an AAV P5 promoter. Optionally, the selected promoter is used in conjunction with a heterologous enhancer, e.g., the β-

actin promoter may be used in conjunction with the CMV enhancer. Yet other suitable or desirable promoters and enhancers may be selected by one of skill in the art.

The minigene may also desirably contain nucleic acid sequences heterologous to the viral vector sequences including sequences providing signals required for efficient polyadenylation of the transcript (poly-A or pA) and introns with functional splice donor and acceptor sites. A common poly-A sequence which is employed in the exemplary vectors of this invention is that derived from the papovavirus SV-40. The poly-A sequence generally is inserted in the minigene downstream of the transgene sequences and upstream of the viral vector sequences. A common intron sequence is also derived from SV-40, and is referred to as the SV40 T intron sequence. A minigene of the present invention may also contain such an intron, desirably located between the promoter/enhancer sequence and the transgene. Selection of these and other common vector elements are conventional [see, e.g., Sambrook et al, "Molecular Cloning. A Laboratory Manual", 2d edit., Cold Spring Harbor Laboratory. New York (1989) and references cited therein] and many such sequences are available from commercial and industrial sources as well as from Genebank.

The selection of the transgene is not a limitation of the present invention. Suitable transgenes may be readily selected from among desirable reporter genes, therapeutic genes, and optionally, genes encoding immunogenic polypeptides. Examples of suitable reporter genes include β -galactosidase (β -gal), an alkaline phosphatase gene, and green fluorescent protein (GFP). Examples of therapeutic genes include, cytokines, growth factors, hormones, and differentiation factors, among others. The transgene may be readily selected by one of skill in the art. See, e.g., WO 98/09657, which identifies other suitable transgenes.

Suitably, the vectors of the invention contain, at a minimum, cassettes which consist of fragments of the AAV-1 sequences and proteins. In one embodiment, a vector of the invention comprises a selected transgene, which is flanked by a 5' ITR and a 3' ITR, at least one of which is an AAV-1 ITR of the invention. Suitably,

vectors of the invention may contain a AAV-1 P5 promoter of the invention. In yet another embodiment, a plasmid or vector of the invention contains AAV-1 rep sequences. In still another embodiment, a plasmid or vector of the invention contains at least one of the AAV-1 cap proteins of the invention. Most suitably, these AAV-1-derived vectors are assembled into viral vectors, as described herein.

A. AAV Viral Vectors

In one aspect, the present invention provides a recombinant AAV-1 viral vector produced using the AAV-1 capsid proteins of the invention. The packaged rAAV-1 virions of the invention may contain, in addition to a selected minigene, other AAV-1 sequences, or may contain sequences from other AAV serotypes.

Methods of generating rAAV virions are well known and the selection of a suitable method is not a limitation on the present invention. See, e.g., K. Fisher et al, J. Virol., 70:520-532 (1993) and US Patent 5,478,745. In one suitable method, a selected host cell is provided with the AAV sequence encoding a rep protein, the gene encoding the AAV cap protein and with the sequences for packaging and subsequent delivery. Desirably, the method utilizes the sequences encoding the AAV-1 rep and/or cap proteins of the invention.

In one embodiment, the rep/cap genes and the sequences for delivery are supplied by co-transfection of vectors carrying these genes and sequences. In one currently preferred embodiment, a cis (vector) plasmid, a trans plasmid containing the rep and cap genes, and a plasmid containing the adenovirus helper genes are co-transfected into a suitable cell line, e.g., 293. Alternatively, one or more of these functions may be provided in trans via separate vectors, or may be found in a suitably

25 engineered packaging cell line.

An exemplary cis plasmid will contain, in 5' to 3' order, AAV 5' ITR, the selected transgene, and AAV 3' ITR. In one desirable embodiment, at least one of the AAV ITRs is a 143 nt AAV-1 ITR. However, other AAV serotype ITRs may be readily selected. Suitably, the full-length ITRs are utilized. However, one of skill in

the art can readily prepare modified AAV ITRs using conventional techniques.

Similarly, methods for construction of such plasmids is well known to those of skill in the art.

A trans plasmid for use in the production of the rAAV-1 virion particle may be prepared according to known techniques. In one desired embodiment, this plasmid contains the rep and cap proteins of AAV-1, or functional fragments thereof. Alternatively, the rep sequences may be from another selected AAV serotype.

The cis and trans plasmid may then be co-transfected with a wild-type helper virus (e.g., Ad2, Ad5, or a herpesvirus), or more desirably, a replication - defective adenovirus, into a selected host cell. Alternatively, the cis and trans plasmid may be co-transfected into a selected host cell together with a transfected plasmid which provides the necessary helper functions. Selection of a suitable host cell is well within the skill of those in the art and include such mammalian cells as 293 cells, HeLa cells, among others.

Alternatively, the cis plasmid and, optionally the trans plasmid, may be transfected into a packaging cell line which provides the remaining helper functions necessary for production of a rAAV containing the desired AAV-1 sequences of the invention. An example of a suitable packaging cell line, where an AAV-2 capsid is desired, is B-50, which stably expresses AAV-2 rep and cap genes under the control of a homologous P5 promoter. This cell line is characterized by integration into the cellular chromosome of multiple copies (at least 5 copies) of P5-rep-cap gene cassettes in a concatomer form. This B-50 cell line was deposited with the American Type Culture Collection, 10801 University Boulevard, Manassas, Virginia 20110-2209, on September 18, 1997 under Accession No. CRL-12401 pursuant to the provisions of the Budapest Treaty. However, the present invention is not limited as to the selection of the packaging cell line.

Exemplary transducing vectors based on AAV-1 capsid proteins have been tested both *in vivo and in vitro*, as described in more detail in Example 4. In these studies, it was demonstrated that recombinant AAV vector with an AAV-1 virion can transduce both mouse liver and muscle. These, and other AAV-1 based

gene therapy vectors which may be generated by one of skill in the art are beneficial for gene delivery to selected host cells and gene therapy patients since the neutralization antibodies of AAV-1 present in much of the human population exhibit different patterns from other AAV serotypes and therefore do not neutralize the AAV-1 virions. One of skill in the art may readily prepare other rAAV viral vectors containing the AAV-1 capsid proteins provided herein using a variety of techniques known to those of skill in the art. One may similarly prepare still other rAAV viral vectors containing AAV-1 sequence and AAV capsids of another serotype.

B. Other Viral Vectors

One of skill in the art will readily understand that the AAV-1 sequences of the invention can be readily adapted for use in these and other viral vector systems for *in vitro*, *ex vivo* or *in vivo* gene delivery. Particularly well suited for use in such viral vector systems are the AAV-1 ITR sequences, the AAV-1 rep, the AAV-1 cap, and the AAV-1 P5 promoter sequences.

For example, in one desirable embodiment, the AAV-1 ITR sequences of the invention may be used in an expression cassette which includes AAV-1 5' ITR, a non-AAV DNA sequences of interest (e.g., a minigene), and 3' ITR and which lacks functional rep/cap. Such a cassette containing an AAV-1 ITR may be located on a plasmid for subsequent transfection into a desired host cell, such as the cis plasmid described above. This expression cassette may further be provided with an AAV capsid of a selected serotype to permit infection of a cell or stably transfected into a desired host cell for packaging of rAAV virions. Such an expression cassette may be readily adapted for use in other viral systems, including adenovirus systems and lentivirus systems. Methods of producing Ad/AAV vectors are well known to those of skill in the art. One desirable method is described in PCT/US95/14018. However, the present invention is not limited to any particular method.

Another aspect of the present invention is the novel AAV-1 P5 promoter sequences which are located in the region spanning nt 236 - 299 of SEQ ID NO: 1. This promoter is useful in a variety of viral vectors for driving expression of a desired transgene.

Similarly, one of skill in the art can readily select other fragments of the AAV-1 genome of the invention for use in a variety of vector systems. Such vectors systems may include, e.g., lentiviruses, retroviruses, poxviruses, vaccinia viruses, and adenoviral systems, among others. Selection of these vector systems is not a limitation of the present invention.

C. Host Cells And Packaging Cell Lines

In yet another aspect, the present invention provides host cells which may be transfected with AAV-1 nucleic acid sequences of the invention to permit expression of a desired transgene or production of a rAAV particle. For example, a selected host cell may be transfected with the AAV-1 P5 promoter sequences and/or the AAV-1 5' ITR sequences using conventional techniques.

Providing AAV helper functions to the transfected cell lines of the invention results in packaging of the rAAV as infectious rAAV particles. Such cell lines may be produced in accordance with known techniques [see, e.g, US Patent No. 5,658,785], making use of the AAV-1 sequences of the invention.

Alternatively, host cells of the invention may be stably transfected with a rAAV expression cassette of the invention, and with copies of AAV-1 rep and cap genes. Suitable parental cell lines include mammalian cell lines and it may be desirable to select host cells from among non-simian mammalian cells. Examples of suitable parental cell lines include, without limitation, HeLa [ATCC CCL 2], A549 [ATCC Accession No. CCL 185], KB [CCL 17], Detroit [e.g., Detroit 510, CCL 72] and WI-38 [CCL 75] cells. These cell lines are all available from the American Type Culture Collection, 10801 University Boulevard, Manassas, Virginia 20110-2209 USA. Other suitable parent cell lines may be obtained from other sources and may be used to construct stable cell lines containing the P5 and/or AAV rep and cap sequences of the invention.

Recombinant vectors generated as described above are useful for delivery of the DNA of interest to cells.

III. METHODS OF DELIVERING GENES VIA AAV-1 DERIVED VECTORS

In another aspect, the present invention provides a method for delivery of a transgene to a host which involves transfecting or infecting a selected host cell with a recombinant viral vector generated with the AAV-1 sequences (or functional fragments thereof) of the invention. Methods for delivery are well known to those of skill in the art and are not a limitation of the present invention.

In one desirable embodiment, the invention provides a method for AAV-mediated delivery of a transgene to a host. This method involves transfecting or
infecting a selected host cell with a recombinant viral vector containing a selected
transgene under the control of sequences which direct expression thereof and AAV-1
capsid proteins.

Optionally, a sample from the host may be first assayed for the presence of antibodies to a selected AAV serotype. A variety of assay formats for detecting neutralizing antibodies are well known to those of skill in the art. The selection of such an assay is not a limitation of the present invention. See, e.g., Fisher et al, Nature Med., 3(3):306-312 (March 1997) and W. C. Manning et al, Human Gene Therapy, 2:477-485 (March 1, 1998). The results of this assay may be used to determine which AAV vector containing capsid proteins of a particular serotype are preferred for delivery, e.g., by the absence of neutralizing antibodies specific for that capsid serotype.

In one aspect of this method, the delivery of vector with AAV-1 capsid proteins may precede or follow delivery of a gene via a vector with a different serotype AAV capsid protein. Thus, gene delivery via rAAV vectors may be used for repeat gene delivery to a selected host cell. Desirably, subsequently administered rAAV vectors carry the same transgene as the first rAAV vector, but the subsequently administered vectors contain capsid proteins of serotypes which differ from the first vector. For example, if a first vector has AAV-2 capsid proteins, subsequently administered vectors may have capsid proteins selected from among the other serotypes, including AAV-1, AAV-3A, AAV-3B, AAV-4 and AAV-6.

Thus, a rAAV-1-derived recombinant viral vector of the invention provides an efficient gene transfer vehicle which can deliver a selected transgene to a selected host cell *in vivo or ex vivo* even where the organism has neutralizing antibodies to one or more AAV serotypes. These compositions are particularly well suited to gene delivery for therapeutic purposes. However, the compositions of the invention may also be useful in immunization. Further, the compositions of the invention may also be used for production of a desired gene product *in vitro*.

The above-described recombinant vectors may be delivered to host cells according to published methods. An AAV viral vector bearing the selected transgene may be administered to a patient, preferably suspended in a biologically compatible solution or pharmaceutically acceptable delivery vehicle. A suitable vehicle includes sterile saline. Other aqueous and non-aqueous isotonic sterile injection solutions and aqueous and non-aqueous sterile suspensions known to be pharmaceutically acceptable carriers and well known to those of skill in the art may be employed for this purpose.

The viral vectors are administered in sufficient amounts to transfect the cells and to provide sufficient levels of gene transfer and expression to provide a therapeutic benefit without undue adverse effects, or with medically acceptable physiological effects, which can be determined by those skilled in the medical arts. Conventional and pharmaceutically acceptable routes of administration include, but are not limited to, direct delivery to the liver, oral, intranasal, intravenous, intramuscular, subcutaneous, intradermal, and other parental routes of administration. Routes of administration may be combined, if desired.

Dosages of the viral vector will depend primarily on factors such as the condition being treated, the age, weight and health of the patient, and may thus vary among patients. For example, a therapeutically effective human dosage of the viral vector is generally in the range of from about 1 ml to about 100 ml of solution containing concentrations of from about 1×10^9 to 1×10^{16} genomes virus vector. A preferred human dosage may be about 1×10^{13} to 1×10^{16} AAV genomes. The dosage will be adjusted to balance the therapeutic benefit against any side effects and

such dosages may vary depending upon the therapeutic application for which the recombinant vector is employed. The levels of expression of the transgene can be monitored to determine the frequency of dosage resulting in viral vectors, preferably AAV vectors containing the minigene. Optionally, dosage regimens similar to those described for therapeutic purposes may be utilized for immunization using the compositions of the invention. For *in vitro* production, a desired protein may be obtained from a desired culture following transfection of host cells with a rAAV containing the gene encoding the desired protein and culturing the cell culture under conditions which permits expression. The expressed protein may then be purified and isolated, as desired. Suitable techniques for transfection, cell culturing, purification, and isolation are known to those of skill in the art.

The following examples illustrate several aspects and embodiments of the invention.

Example 1 - Generation of Infectious Clone of AAV-1

The replicated form DNA of AAV-1 was extracted from 293 cells that were infected by AAV-1 and wild type adenovirus type 5.

A. Cell Culture and Virus

AAV-free 293 cells and 84-31 cells were provided by the human application laboratory of the University of Pennsylvania. These cells were cultured in Dulbecco's Modified Eagle Medium with 10% fetal bovine serum (Hyclone), penicillin (100 U/ml) and streptomycin at 37°C in a moisturized environment supplied with 5% CO₂. The 84-31 cell line constitutively expresses adenovirus genes E1a, Elb, E4/ORF6, and has been described previously [K. J. Fisher, <u>J. Virol.</u>, <u>70</u>:520-532 (1996)]. AAV-1 (ATCC VR-645) seed stock was purchased from American Type Culture Collection (ATCC, Manassas, VA). AAV viruses were propagated in 293 cells with wild type Ad5 as a helper virus.

B. Recombinant AAV Generation

The recombinant AAV viruses were generated by transfection using an adenovirus free method. Briefly, the cis plasmid (with AAV ITR), trans plasmid (with

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AAV rep gene and cap gene) and helper plasmid (pFa13, with essential regions from the adenovirus genome) were simultaneously co-transfected into 293 cells in a ratio of 1:1:2 by calcium phosphate precipitation. The pFa13 helper plasmid has an 8 kb deletion in the adenovirus E2B region and has deletions in most of the late genes. This helper plasmid was generated by deleting the RsrII fragment from pFG140 (Microbix, Canada). Typically, 50 µg of DNA (cis:trans:PFa13 at ratios of 1:1:2, respectively) was transfected onto a 15 cm tissue culture dish. The cells were harvested 96 hours post-transfection, sonicated and treated with 0.5% sodium deoxycholate (37°C for 10 min). Cell lysates were then subjected to two rounds of a CsCl gradient. Peak fractions containing AAV vector were collected, pooled, and dialyzed against PBS before injecting into animals. To make rAAV virus with AAV-1 virion, the pAV1H or p5E18 (2/1) was used as the *trans* plasmid to provide rep and cap function.

For the generation of rAAV based on AAV-2, p5E18 was used as the trans plasmid since it greatly improved the rAAV yield. This plasmid, p5E18(2/2), expresses AAV-2 Rep and Cap and contains a P5 promoter relocated to a position 3' to the Cap gene, thereby minimizing expression of Rep78 and Rep68. The strategy was initially described by Li et al, J. Virol., 71:5236-5243 (1997). P5E18(2/2) was constructed in the following way. The previously described pMMTV-trans vector (i.e., the mouse mammary tumor virus promoter substituted for the P5 promoter in an AAV-2-based vector) was digested with Smal and ClaI, filled in with the Klenow enzyme, and then recircularized with DNA ligase. The resulting construct was digested with XbaI, filled in, and ligated to the blunt-ended BamHI-XbaI fragment from pCR-p5, constructed in the following way. The P5 promoter of AAV was amplified by PCR and the amplified fragment was subsequently cloned into pCR2.1 (Invitrogen) to yield pCR-P5. The helper plasmid pAV1H was constructed by cloning the BfaI fragment of pAAV-2 into pBluescript II-SK(+) at the BcorV and SmaI sites. The 3.0-kb XbaI-KpnI fragment from p5E18(2/2), the 2.3-kb XbaI-KpnI fragment from pAV1H, and the 1.7-kb KpnI fragment from p5E18(2/2) were incorporated into a separate plasmid P5E18(2/1), which contains AAV-2 Rep, AAV-1 Cap, and the

AAV-2 P5 promoter located 3' to the Cap gene. Plasmid p5E18(2/1) produced 10- to 20-fold higher quantities of the vector than pAV1H (i.e., 10¹² genomes/50 15-cm² plates).

C. DNA Techniques

EcoRV site.

Hirt DNA extraction was performed as described in the art with minor modification [R.J. Samulski et al., Cell, 33:135-143 (1983)]. More particularly, Hirst solution without SDS was used instead of using original Hirt solution containing SDS. The amount of SDS present in the original Hirst solution was added after the cells had been fully suspended. To construct AAV-1 infectious clone, the Hirt DNA from AAV-1 infected 293 cells was repaired with Klenow enzyme (New England Biolabs) to ensure the ends were blunt. The treated AAV-1 Hirt DNA was then digested with BamHI and cloned into three vectors, respectively. The internal BamHI was cloned into pBlueScript II-SK+ cut with BamHI to get pAV1-BM. The left and right fragments were cloned into pBlueScript II-SK+ cut with BamHI + EcoRV to obtain pAV1-BL and pAV1-BR, respectively. The AAV sequence in these three plasmids were subsequently assembled into the same vector to get AAV-1 infectious clone pAAV-1. The helper plasmid for recombinant AAV-1 virus generation was constructed by cloning the Bfa I fragment of pAAV-1 into pBlueScript II-SK+ at the

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Analysis of the Hirt DNA revealed three bands, a dimer at 9.4 kb, a monomer at 4.7 kb and single-stranded DNA at 1.7 kb, which correlated to different replication forms of AAV-1. The monomer band was excised from the gel and then digested with *BamH*I. This resulted in three fragments of 1.1 kb, 0.8 kb and 2.8 kb. This pattern is in accordance with the description by Bantel-schaal and zur Hausen, Virol., 134(1):52-63 (1984). The 1.1 kb and 2.8 kb *BamH*I fragments were cloned into pBlueScript-KS(+) at *BamH*I and EcoRV site. The internal 0.8 kb fragment was cloned into *BamH*I site of pBlueScript-KS(+).

These three fragments were then subcloned into the same construct to obtain a plasmid (pAAV-1) that contained the full sequence of AAV-1. The pAAV-1 was then tested for its ability to rescue from the plasmid backbone and package

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infectious virus. The pAAV-1 was then transfected to 293 cells and supplied with adenovirus type as helper at MOI 10. The virus supernatant was used to reinfect 293 cells.

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For Southern blot analysis, Hirt DNA was digested with *DpnI* to remove bacteria-borne plasmid and probed with internal *BamHI* fragment of AAV-1. The membrane was then washed at high stringency conditions, which included: twice 30 minutes with 2X SSC, 0.1% SDS at 65°C and twice 30 minutes with 0.1X SSC, 0.1% SDS at 65°C. The membrane was then analyzed by both phosphor image and X-ray autoradiography. The results confirmed that pAAV-1 is indeed an infectious clone of AAV serotype 1.

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Example 2 - Sequencing Analysis of AAV-1

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The entire AAV-1 genome was then determined by automatic sequencing and was found to be 4718 nucleotides in length (Figs. 1A-1C). For sequencing, an ABI 373 automatic sequencer as used to determine the sequences for all plasmids and PCR fragments related to this study using the FS dye chemistry. All sequences were confirmed by sequencing both plus and minus strands. These sequences were also confirmed by sequencing two independent clones of pAV-BM, pAV-BL and pAV-BR. Since the replicated form of AAV-1 DNA served as the template for sequence determination, these sequences were also confirmed by sequencing a series of PCR products using original AAV-1 seed stock as a template.

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The length of AAV-1 was found to be within the range of the other serotypes: AAV-3 (4726 nucleotides), AAV-4 (4774 nucleotides), AAV-2 (4681 nucleotides),

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and AAV-6 (4683 nucleotides).

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The AAV-1 genome exhibited similarities to other serotypes of adenoassociated viruses. Overall, it shares more than 80% identity with other known AAV viruses as determined by the computer program Megalign using default settings [DNASTAR, Madison, WI]. The key features in AAV-2 can also be found in AAV-1. First, AAV-1 has the same type of inverted terminal repeat which is capable of

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forming T-shaped hairpin structures, despite the differences at the nucleotide level

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(Figs. 2 and 3). The sequences of right ITRs and left ITRs of AAV-1 are identical. The AAV TR sequence is subdivided into A, A', B, B', C, C', D and D' [Bern, cited above].

These AAV ITR sequences are also virtually the same as those found in AAV-

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6 right ITR, there being one nucleotide difference in each of A and A' sequence, and the last nucleotide of the D sequence. Second, the AAV-2 rep binding motif [GCTCGCTCGCTCGCTG (SEQ ID NO: 20)] is well conserved. Such motif can also be found in the human chromosome 19 AAV-2 pre-integration region. Finally, non-structural and structural coding regions, and regulatory elements similar to those

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of other AAV serotypes also exist in AAV-1 genome.

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conserved, the total length of the AAV terminal repeat exhibits divergence. The terminal repeat of AAV-1 consists of 143 nucleotides while those of AAV-2, AAV-3, and AAV-4 are about 145 or 146 nucleotides. The loop region of AAV-1 ITR most

Although the overall features of AAV terminal repeats are very much

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closely resembles that of AAV-4 in that it also uses TCT instead of the TTT found in

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AAV-2 and AAV-3. The possibility of sequencing error was eliminated using restriction enzyme digestion, since these three nucleotides are part of the SacI site (gagete; nt 69-74 of SEQ ID NO: 1). The p5 promoter region of AAV-1 shows more

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maintains the critical regulatory elements. The two copies of YY1 [See, Fig. 1A-1C] sites seemed to be preserved in all known AAV serotypes, which have been shown to

variations in nucleotide sequences with other AAV serotypes. However, it still

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be involved in regulating AAV gene expression. In AAV-4, there are 56 additional nucleotides inserted between YY1 and E-box/USF site, while in AAV-1, there are 26

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promoter and polyA can also be identified from the AAV-1 genome by analogy to known AAV serotypes, which are also highly conserved.

Thus, the analysis of AAV terminal repeats of various serotypes showed that

additional nucleotides inserted before the E-box/USF site. The p19 promoter, p40

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the A and A' sequence is very much conserved. One of the reasons may be the Rep binding motif (GCTC)₃GCTG [SEQ ID NO: 20]. These sequences appear to be essential for AAV DNA replication and site-specific integration. The same sequence

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has also been shown to be preserved in a monkey genome [Samulski, personal communication]. The first 8 nucleotides of the D sequence are also identical in all known AAV serotypes. This is in accordance with the observation of the Srivastava group that only the first 10 nucleotides are essential for AAV packaging [X.S. Wang et al, J. Virol., 71:3077-3082 (1997); X.S. Wang et al, J. Virol., 71:1140-1146 (1997)]. The function of the rest of the D sequences still remain unclear. They may be somehow related to their tissue specificities. The variation of nucleotide in B and C sequence may also suggest that the secondary structure of the ITRs is more critical for its biological function, which has been demonstrated in many previous publications.

Example 3 - Comparison of AAV-1 Sequences

The nucleotide sequences of AAV-1, obtained as described above, were compared with known AAV sequences, including AAV-2, AAV-4 and AAV-6 using DNA Star Megalign. This comparison revealed a stretch of 71 identical nucleotides shared by AAV-1, AAV-2 and AAV-6. See, Figs. 1A-1C.

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This comparison further suggested that AAV-6 is a hybrid formed by homologous recombination of AAV-1 and AAV-2. See, Figs. 3A and 3B. These nucleotides divide the AAV-6 genome into two regions. The 5' half of AAV-6 of 522 nucleotides is identical to that of AAV-2 except in 2 positions. The 3' half of AAV-6 including the majority of the rep gene, complete cap gene and 3' ITR is 98% identical to AAV-1.

Biologically, such recombination may enable AAV-1 to acquire the ability to transmit through the human population. It is also interesting to note that the ITRs of AAV-6 comprise one AAV-1 ITR and one AAV-2 ITR. The replication model of defective parvovirus can maintain this special arrangement. Studies on AAV integration have shown that a majority of AAV integrants carries deletions in at least one of the terminal repeats. These deletions have been shown to be able to be repaired through gene conversion using the other intact terminal repeat as a template. Therefore, it would be very difficult to maintain AAV-6 as a homogenous population

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when an integrated copy of AAV-6 is rescued from host cells with helper virus infection. The AAV-6 with two identical AAV-2 ITRs or two identical AAV-1 ITRs should be the dominant variants. The AAV-6 with two AAV-1 ITRs has been observed by Russell's group [Rutledge, cited above (1998)]. So far there is no report on AAV-6 with two AAV-2 ITRs. Acquirement of AAV-2 P5 promoter by AAV-6 may have explained that AAV-6 have been isolated from human origin while AAV-1 with the same virion has not. The regulation of P5 promoter between different species of AAV may be different in vivo. This observation suggests the capsid proteins of AAV were not the only determinants for tissue specificity.

Although it is clear that AAV-6 is a hybrid of AAV-1 and AAV-2, AAV-6 has already exhibited divergence from either AAV-1 or AAV-2. There are two nucleotide differences between AAV-6 and AAV-2 in their first 450 nucleotides. There are about 1% differences between AAV-6 and AAV-1 in nucleotide levels from nucleotides 522 to the 3' end. There also exists a quite divergent region (nucleotide 4486-4593) between AAV-6 and AAV-1 (Figs. 1A-1C). This region does not encode any known proteins for AAVs. These differences in nucleotide sequences may suggest that AAV-6 and AAV-1 have gone through some evolution since the recombination took place. Another possible explanation is that there exists another variant of AAV-1 which has yet to be identified. So far, there is no evidence to rule out either possibility. It is still unknown if other hybrids (AAV-2 to AAV-4, etc.)

existed in nature.

The coding region of AAV-1 was deduced by comparison with other known AAV serotypes. Table 1 illustrates the coding region differences between AAV-1 and AAV-6. The amino acid residues are deduced according to AAV-2.

With reference to the amino acid position of AAV-1, Table 1 lists the amino acids of AAV-1 which have been changed to the corresponding ones of AAV-6. The amino acids of AAV-1 are shown to the left of the arrow. Reference may be made to SEQ ID NO: 5 of the amino acid sequence of AAV-1 Rep 78 and to SEQ ID NO: 13 for the amino acid sequence of AAV-1 VP1.

Table 1
Coding region variations between AAV-1 and AAV-6

Rep prote	ein (Rep78)	Cap pro	Cap protein (VP1)		
Position(s) Amino acids		Position(s)	Amino acids		
28	S-N	129	L→F		
191	Q~H	418	E-D		
192	H-D	531	E-K		
308	E-D	584	F-L		
		598	A-V		
		642	N−H		

It was surprising to see that the sequence of the AAV-1 coding region is almost identical to that of AAV-6 from position 452 to the end of coding region (99%). The first 508 nucleotides of AAV-6 have been shown to be identical to those of AAV-2 [Rutledge, cited above (1998)]. Since the components of AAV-6 genome seemed to be AAV-2 left ITR - AAV-2 p5 promoter - AAV-1 coding region - AAV-1 right ITR, it was concluded that AAV-6 is a naturally occurred hybrid between AAV-1 and AAV-2.

Example 4 - Gene Therapy Vector Based on AAV-1

Recombinant gene transfer vectors based on AAV-1 viruses were constructed by the methods described in Example 1. To produce a hybrid recombinant virus with AAV-1 virion and AAV-2 ITR, the AAV-1 trans plasmid (pAV1H) and the AAV-2 cis-lacZ plasmid (with AAV-2 ITR) were used. The AAV-2 ITR was used in this vector in view of its known ability to direct site-specific integration. Also constructed for use in this experiment was an AAV-1 vector carrying the green fluorescent protein (GFP) marker gene under the control of the immediate early promoter of CMV using pAV1H as the trans plasmid.

A. rAAV-1 Viruses Transfect Host Cells in Vitro

84-31 cells, which are subclones of 293 cells (which express adenovirus E1a, E1b) which stably express E4/ORF5, were infected with rAAV-1 GFP or rAAV-lacZ. High levels of expression of GFP and lacZ was detected in the cultured 84-31 cells. This suggested that rAAV-1 based vector was very similar to AAV-2 based vectors in ability to infect and expression levels.

B. rAAV-1 Viruses Transfect Cells in Vivo

The performance of AAV-1 based vectors was also tested *in vivo*. The rAAV-1 CMV-α1AT virus was constructed as follows. The EcoRI fragment of pAT85 (ATCC) containing human α1-antitrypsin (α1AT) cDNA fragment was blunted and cloned into PCR (Promega) at a Smal site to obtain PCR-α1AT. The CMV promoter was cloned into PCR-α1AT at the Xbal site. The Alb-α1AT expression cassette was removed by XhoI and ClaI and cloned into pAV1H at the Xbal site. This vector plasmid was used to generate AAV-1-CMV-α1AT virus used in the experiment described below.

For screening human antibodies against AAV, purified AAV virus is lysed with Ripa buffer (10 mM Tris pH 8.2, 1% Triton X-100, 1% SDS, 0.15 M NaCl) and separated in 10% SDS-PAGE gel. The heat inactivated human serum was used at a 1 to 1000 dilution in this assay. The rAAV-1 CMV- α 1AT viruses were injected into Rag-1 mice through tail vein injection at different dosages. The concentration of human α 1-antitrypsin in mouse serum was measured using ELISA. The coating antibody is rabbit anti-human human α 1-antitrypsin (Sigma). The goatantihuman α 1-antitrypsin (Sigma) was used as the primary detection antibodies. The sensitivity of this assay is around 0.3 ng/ml to 30 ng/ml. The expression of human α -antitrypsin in mouse blood can be detected in a very encouraging level. This result is shown in Table 2.

Table 2 Human Antitrypsin Expressed in Mouse Liver

Amount of virus injected	Week 2 (ng/ml)	Week 4 (ng/ml)
2x10 ¹⁰ genomes	214.2	171.4
1x10 ¹⁰ genomes	117.8	109.8
5x10 ¹⁰ genomes	64.5	67.8
2.5x10 ¹⁰ genomes	30.9	58.4

rAAV-1 CMV-lacZ viruses were also injected into the muscle of

C57BL6 mice and similar results were obtained. Collectively, these results suggested
that AAV-1 based vector would be appropriate for both liver and muscle gene
delivery.

Example 5 - Neutralizing Antibodies Against AAV-1

Simple and quantitative assays for neutralizing antibodies (NAB) to AAV-1 and AAV-2 were developed with recombinant vectors. A total of 33 rhesus monkeys and 77 normal human subjects were screened.

A. Nonhuman Primates

Wild-caught juvenile rhesus monkeys were purchased from Covance (Alice, Tex.) and LABS of Virginia (Yemassee, SC) and kept in full quarantine. The monkeys weighed approximately 3 to 4 kg. The nonhuman primates used in the Institute for Human Gene Therapy research program are purposefully bred in the United States from specific-pathogen-free closed colonies. All vendors are US Department of Agriculture class A dealers. The rhesus macaques are therefore not infected with important simian pathogens, including the tuberculosis agent, major simian lentiviruses (simian immunodeficiency virus and simian retroviruses), and cercopithecine herpesvirus. The animals are also free of internal and external parasites. The excellent health status of these premium animals minimized the potential for extraneous variables. For this study, serum was obtained from monkeys prior to initiation of any protocol.

NAB titers were analyzed by assessing the ability of serum antibody to inhibit the transduction of reporter virus expressing green fluorescent protein (GFP) (AAV1-GFP or AAV2-GFP) into 84-31 cells. Various dilutions of antibodies preincubated with reporter virus for 1 hour at 37°C were added to 90% confluent cell cultures. Cells were incubated for 48 hours and the expression of green fluorescent protein was measured by FluoroImaging (Molecular Dynamics). NAB titers were calculated as the highest dilution at which 50% of the cells stained green.

Analysis of NAB in rhesus monkeys showed that 61% of animals tested positive for AAV-1; a minority (24%) has NAB to AAV-2. Over one-third of animals had antibodies to AAV-1 but not AAV-2 (i.e., were monospecific for AAV-1), whereas no animals were positive for AAV-2 without reacting to AAV-1. These data support the hypothesis that AAV-1 is endemic in rhesus monkeys. The presence of true AAV-2 infections in this group of nonhuman primates is less clear, since crossneutralizing activity of an AAV-1 response to AAV-2 can not be ruled out. It is interesting that there is a linear relationship between AAV-2 NAB and AAV-1 NAB in animals that had both.

B. Humans

For these neutralization antibody assays, human serum samples were incubated at 56°C for 30 min to inactivate complement and then diluted in DMEM. The virus (rAAV or rAd with either lacZ or GFP) was then mixed with each serum dilution (20X, 400X, 2000X, 4000X, etc.) and incubated for 1 hour at 37°C before applied to 90% confluent cultures of 84-31 cells (for AAV) or Hela cells (for adenovirus) in 96-well plates. After 60 minutes of incubation at culture condition, 100 µl additional media containing 20% FCS was added to make final culture media containing 10% FCS.

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The result is summarized in Table 3.

Table 3

Adenovirus	AAV-1	AAV-2	# of samples	Percentage
-	-	-	41	53.2%
+	-	-	16	20.8%
•	+	-	0	0.0%
-	-	+	2	2.6%
•	+	+	2	2.6%
+	-	÷	3	3.9%
+	+	-	0	0.0%
+	+	+	13	16.9%
		Total	77	100%

The human neutralizing antibodies against these three viruses seemed to be unrelated since the existence of neutralizing antibodies against AAV are not indications for antibodies against adenovirus. However, AAV requires adenovirus as helper virus, in most of the cases, the neutralizing antibodies against AAV correlated with the existence of neutralizing antibodies to adenovirus. Among the 77 human serum samples screened, 41% of the samples can neutralize the infectivity of recombinant adenovirus based on Ad5. 15/77 (19%) of serum samples can neutralize the transduction of rAAV-1 while 20/77 (20%) of the samples inhibit rAAV-2 transduction at 1 to 80 dilutions or higher. All serum samples positive in neutralizing antibodies for AAV-1 in are also positive for AAV-2. However, there are five (6%) rAAV-2 positive samples that failed to neutralize rAAV-1. In samples that are positive for neutralizing antibodies, the titer of antibodies also varied in the positive ones. The results from screening human sera for antibodies against AAVs supported the conclusion that AAV-1 presents the same epitome as that of AAV-2 to interact

with cellular receptors since AAV-1 neutralizing human serums can also decrease the infectivity of AAV-2. However, the profile of neutralizing antibodies for these AAVs is not identical, there are additional specific receptors for each AAV serotype.

Example 6 - Recombinant AAV Viruses Exhibit Tissue Tropism

The recombinant AAV-1 vectors of the invention and the recombinant AAV-2 vectors [containing the gene encoding human α 1-antitrypsin (α 1AT) or murine erythropoietin (Epo) from a cytomegalovirus-enhanced β -actin promoter (CB)] were evaluated in a direct comparison to equivalent copies of AAV-2 vectors containing the same vector genes.

Recombinant viruses with AAV-1 capsids were constructed using the techniques in Example 1. To make rAAV with AAV-1 virions, pAV1H or p5E18 (2/1) was used as the *trans* plasmid to provide Rep and Cap functions. For the generation of the rAAV based on AAV-2, p5E18(2/2) was used as the *trans* plasmid, since it greatly improved the rAAV yield. [Early experiments indicated similar *in vivo* performances of AAV-1 vectors produced with pAV1H and p5E19 (2/1). All subsequent studies used AAV-1 vectors derived from p5E18(2/1) because of the increased yield.]

Equivalent stocks of the AAV-1 and AAV-2 vectors were injected intramuscularly (5 x 10¹⁰ genomes) or liver via the portal circulation (1 x 10¹¹ genomes) into immunodeficient mice, and the animals (four groups) were analyzed on day 30 for expression of transgene. See, Figs. 4A and 4B.

AAV-2 vectors consistently produced 10- to 50-fold more serum erythropoietin or a1-antitrypsin when injected into liver compared to muscle. (However, the AAV-1-delivered genes did achieve acceptable expression levels in the liver.) This result was very different from that for AAV-1 vectors, with which muscle expression was equivalent to or greater than liver expression. In fact, AAV-1 outperformed AAV-2 in muscle when equivalent titers based on genomes were administered.

Example 7 - Gene Delivery via rAAV-1

C57BL/6 mice (6- to 8-week old males, Jackson Laboratories) were analyzed for AAV mediated gene transfer to liver following intrasplenic injection of vector (i.e., targeted to liver). A total of 10¹¹ genome equivalents of rAAV-1 or rAAV-2 vector were injected into the circulation in 100 μl buffered saline. The first vector contained either an AAV-1 capsid or an AAV-2 capsid and expressed α1AT under the control of the chicken β-actin (CB) promoter. Day 28 sera were analyzed for antibodies against AAV-1 or AAV-2 and serum α1AT levels were checked. Animals were then injected with an AAV-1 or AAV-2 construct expressing erythropoietin (Epo, also under the control of the CB promoter). One month later sera was analyzed for serum levels of Epo. The following groups were analyzed (Figs. 5A-5D).

In Group 1, vector 1 was AAV-2 expressing a1AT and vector 2 was AAV-2 expressing Epo. Animals generated antibodies against AAV-2 following the first vector administration which prevented the readministration of the AAV-2 based vector. There was no evidence for cross-neutralizing the antibody to AAV-1.

In Group 2, vector 1 was AAV-1 expressing a1AT while vector 2 was AAV-1 expressing Epo. The first vector administration did result in significant a1AT expression at one month associated with antibodies to neutralizing antibodies to AAV-1. The animals were not successfully readministered with the AAV-1 Epo expressing construct.

In Group 3, the effectiveness of an AAV-2 vector expressing Epo injected into a naive animal was measured. The animals were injected with PBS and injected with AAV-2 Epo vector at day 28 and analyzed for Epo expression one month later. The neutralizing antibodies were evaluated at day 28 so we did not expect to see anything since they received PBS with the first vector injection. This shows that in naive animals AAV-2 is very efficient at transferring the Epo gene as demonstrated by high level of serum Epo one month later.

Group 4 was an experiment similar to Group 3 in which the animals originally received PBS for vector 1 and then the AAV-1 expressing Epo construct 28 days later. At the time of vector injection, there obviously were no antibodies to either

AAV-1 or AAV-2. The AAV-1 based vector was capable of generating significant expression of Epo when measured one month later.

Group 5 is a cross-over experiment where the initial vector is AAV-2 expressing α1AT followed by the AAV-1 construct expressing Epo. The animals, as expected, were efficiently infected with the AAV-2 vector expressing α1AT as shown by high levels of the protein in blood at 28 days. This was associated with significant neutralizing antibodies to AAV-2. Importantly, the animals were successfully administered AAV-1 following the AAV-2 vector as shown by the presence of Epo in serum 28 days following the second vector administration. At the time of this vector administration, there was high level AAV-2 neutralizing antibodies and very low cross-reaction to AAV-1. The level of Epo was slightly diminished possibly due to a small amount of cross-reactivity. Group 6 was the opposite cross-over experiment in which the initial vector was AAV-1 based, whereas the second experiment was AAV-2 based. The AAV-1 vector did lead to significant gene expression of α1AT, which also resulted in high level AAV-1 neutralizing antibody. The animals were very efficiently administered AAV-2 following the initial AAV-1 vector as evidenced by high level Epo.

A substantially identical experiment was performed in muscle in which 5 x 10¹⁰ genomes were injected into the tibialis anterior of C57BL/6 mice as a model for muscle directed gene therapy. The results are illustrated in Figs. 6A-6D and are essentially the same as for liver.

In summary, this experiment demonstrates the utility of using an AAV-1 vector in patients who have pre-existing antibodies to AAV-2 or who had initially received an AAV-2 vector and need readministration.

25 Example 8 - Construction of Recombinant Viruses Containing AAV-1 ITRs

This example illustrates the construction of recombinant AAV vectors which contain AAV-1 ITRs of the invention.

An AAV-1 cis plasmid is constructed as follows. A 160 bp Xho-NruI AAV-1 fragment containing the AAV-1 5' ITR is obtained from pAV1-BL. pAV1-BL was

generated as described in Example 1. The Xho-NruI fragment is then cloned into a second pAV1-BL plasmid at an XbaI site to provide the plasmid with two AAV-1 ITRs. The desired transgene is then cloned into the modified pAV-1BL at the NruI and BamHI site, which is located between the AAV-1 ITR sequences. The resulting AAV-1 cis plasmid contains AAV-1 ITRs flanking the transgene and lacks functional AAV-1 rep and cap.

Recombinant AAV is produced by simultaneously transfecting three plasmids into 293 cells. These include the AAV-1 cis plasmid described above; a trans plasmid which provides AAV rep/cap functions and lacks AAV ITRs; and a plasmid providing adenovirus helper functions. The rep and/or cap functions may be provided in trans by AAV-1 or another AAV serotype, depending on the immunity profile of the intended recipient. Alternatively, the rep or cap functions may be provided in cis by AAV-1 or another serotype, again depending on the patient's immunity profile.

In a typical cotransfection, 50 µg of DNA (cis:trans:helper at ratios of 1:1:2, respectively) is transfected onto a 15 cm tissue culture dish. Cells are harvested 96 hours post transfection, sonicated and treated with 0.5% sodium deoxycholate (37° for 10 min). Cell lysates are then subjected to 2-3 rounds of ultracentrifugation in a cesium gradient. Peak fractions containing rAAV are collected, pooled and dialyzed against PBS. A typical yield is 1 x 10¹³ genomes/10⁹ cells.

Using this method, one recombinant virus construct is prepared which contains the AAV-1 ITRs flanking the transgene, with an AAV-1 capsid. Another recombinant virus construct is prepared with contains the AAV-1 ITRs flanking the transgene, with an AAV-2 capsid.

All publications cited in this specification are incorporated herein by reference. While the invention has been described with reference to a particularly preferred embodiments, it will be appreciated that modifications can be made without departing from the spirit of the invention. Such modifications are intended to fall within the scope of the claims.

Claims

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			33
	What is claimed	d is:	
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	1.	An isol	ated AAV-1 nucleic acid molecule comprising a sequence
	selected from th	he grou	p consisting of:
15	((a)	SEQ ID NO: 1;
15	((b)	a DNA sequence complementary to SEQ ID NO: 1;
	((c)	cDNA complementary to (a) or (b); and
	((d)	RNA complementary to any of (a) to (c).
20			
	2.	A nucle	eic acid molecule comprising an AAV-1 inverted terminal repeat
	(ITR) sequence	selecte	ed from the group consisting of:
25	((a)	nt 1 to 143 of SEQ ID NO: 1;
23	((b)	nt 4576 to 4718 of SEQ ID NO: 1;
	((c)	a nucleic acid sequence complementary to (a) or (b); and
	((d)	a functional fragment of (a), (b), or (c).
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	3. A	A recon	nbinant vector comprising a 5' AAV-1 inverted terminal repeat
	(ITR) and a sele	ected tra	ansgene, wherein said ITR has the sequence selected from the
35	group consisting	g of:	
	(;	(a) i	nt 1 to 143 of SEQ ID NO: 1;
	(1	(b) a	a nucleic acid sequence complementary to (a); and
	(4	(c) a	a functional fragment of (a) or (b).
40			
	4. T	The rece	ombinant vector according to claim 3, wherein said vector
	further comprise	es a 3' A	AAV-1 ITR.
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		34			
	5 .	A recombinant vector comprising a 3' AAV-1 inverted terminal repeat			
10	(ITR) and a s	selected transgene, wherein said ITR has the sequence selected from the			
	group consist	ting of:			
		(a) nt 4576 to 4718 of SEQ ID NO: 1;			
15		(b) a nucleic acid sequence complementary to (a); and			
75		(c) a functional fragment of (a) or (b).			
	6.	The recombinant vector according to claim 5, wherein said vector			
20	further compr	rises a 5' AAV-1 ITR.			
	7.	The recombinant vector according to any of claims 3-6, wherein said			
25	vector further comprises AAV-1 capsid proteins having the sequence of SEQ ID NO:				
20	13, 15 or 17 o	or functional fragments thereof.			
	8.	The recombinant vector according to any of claims 3-6, wherein said			
30	vector further	r comprises adenovirus sequences.			
	9.	A recombinant vector comprising an AAV-1 P5 promoter having the			
35	sequence of n	at 236 to 299 of SEQ ID NO: 1 or a functional fragment thereof.			
	10.	A nucleic acid molecule encoding AAV-1 helper functions, said			
	molecule com	prising an AAV rep coding region and an AAV cap coding region,			
40	wherein said o	cap coding region comprises at least one member is selected from the			
	group consisti	ing of:			
		(a) vp1, nt 2223 to 4431 of SEQ ID NO: 1;			
45		(b) vp2, nt 2634 to 4432 of SEQ ID NO: 1; and			
		(c) vp3, nt 2829 to 4432 of SEQ ID NO: 1.			

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molecule comprising an AAV rep coding region and an AAV cap coding region,

A nucleic acid molecule encoding AAV-1 helper functions, said

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corresponding thereto;

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wherein said rep coding region comprises an AAV-1 rep coding region comprising at least one member selected from the group consisting of:

(a) rep 78, nt 335 to 2304 of SEQ ID NO: 1;

(b) rep 68, nt 335 to 2272 of SEQ ID NO: 1 or the cDNA

- (c) rep 52, nt 1007 to 2304 of SEQ ID NO: 1; and
- (d) rep 40, nt 1007 to 2272 of SEQ ID NO: 1 or the cDNA corresponding thereto.
- 12. A host cell transduced with a recombinant viral vector according to any of claims 3-6.
- 13. A host cell transduced with a nucleic acid molecule according to any of claims 1, 2, 10 or 11.
- 14. A host cell stably transduced with an AAV-1 P5 promoter having the sequence of nt 236 to 299 of SEQ ID NO: 1.
- 15. A pharmaceutical composition comprising a carrier and a virus comprising the vector according to any of claims 3-6.
- 16. A pharmaceutical composition comprising a carrier and a virus comprising the vector according to claim 7.
- A pharmaceutical composition comprising a carrier and a virus comprising the vector according to claim 8.

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18. A method for AAV-mediated delivery of a transgene comprising the step of delivering to a host cell an AAV virion which comprises:

- (a) a capsid comprising at least one capsid protein encoded by an AAV-1 cap gene; and
- (b) a DNA molecule comprising a transgene under the control of regulatory sequences directing its expression.
- 19. A method for AAV-mediated delivery of a transgene to a host comprising the steps of:
- (a) assaying a sample from the host to determine the presence of neutralizing antibodies specific against any serotype of AAV; and
 - (b) delivering to the host an AAV virion which comprises:
- (i) a capsid comprising at least one capsid protein encoded by a cap gene of an AAV serotype against which the host has no antibodies as determined in step (a); and
- (ii) a DNA molecule comprising a transgene under the control of regulatory sequences directing its expression.
- 20. The method according to claim 19, comprising the additional step of repeating steps (a) and (b).
- 21. Use of an AAV virion which comprises a capsid comprising (a) at least one capsid protein encoded by a cap gene of an AAV serotype against which the host has antibodies, and (b) a DNA molecule comprising a transgene operably linked to regulatory sequences directing its expression,

in the preparation of a medicament for delivery of a transgene to a host, wherein said host has no preexisting neutralizing antibodies against the AAV serotype of said cap gene.

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22. A method for delivery of a transgene comprising the step of delivering to a host cell a recombinant virus comprising a recombinant vector according to any of claims 3-8.

23. A method for producing a selected gene product comprising the steps of transfecting a mammalian cell with the molecule according to claim 1 or a functional fragment thereof and culturing said cell under conditions suitable to express said gene product.

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FIG 1A

AAV-1	ttgcccactccctctgcgcgctcgctcgctcggtggggcctgcggaccaaaggtccgcgg.	60 60
AAV-6	gg.gc	60
	Rep binding site	
AAV-1	agacggcagagctctgctctgccggccdcaccgagcgagcgagcgcgcagagagggagtg	120 120
AAV-2	cc.c.gtc.g.gtgt	120
AAV-6	cc.c.g.,	
7	TRS'	180
AAV-1	ggcaactccatcactaggggtaaTCGCGAAGCGCCTCCCACGCTGCCGCGTCAGCGCTGA .ct.GGTG.AG	163
AAV-2	.cTG.AG	163
	E box/USF	
	CGTAAATTACGTCATAGGGGAGTGGTCCTGTATTAGCTGTCACGTGAGTGCTTTTGC	237
AAV-1	GAGAG	222
AAV-2	GTTA.G.AAG	222
ALV U		
	YY1 P5/TATA	297
AAV-1	GACATTTTGCGACACCACGTGGCCATTTAGGGTATATATGGCCGAGTGAGCAGGAT T. CGCT. T. A.C. ACG.	282
AAV-2	T T CGCT T A.C AC	282
AAV-6		
	YY1/p5 RNA Rep_78/68	356
AAV-1	CTCCATTTTGAC-CGCGAAATTTGAACGAGCAGCCATGCCGGGCTTCTACGAGATCG	342
AAV-2	AGGGGC	341
AAV-1	TGATCAAGGTGCCGAGCGACCTGGACGAGCACCTGCCGGGCATTTCTGACTCGTTTGTGA	416
AAV-2	TCCTGTCAGC	402
	TCC	401
AAV-1	GCTGGGTGGCCGAGAAGGAATGGGAGCTGCCCCCGGATTCTGACATGGATCTGAATCTGA	476
7.77.7	n TG.A	462 461
AAV-6	A	401
AAV-1	TTGAGCAGGCACCCCTGACCGTGGCCGAGAAGCTGCAGCGCGACTTCCTGGTCCAATGGC	536
DDV-2	TACGG	522
AAV-6		521
AAV-1	GCCGCGTGAGTAAGGCCCCGGAGGCCCTCTTCTTTGTTCAGTTCGAGAAGGGCGAGTCCT	596
DAV-2	TAAG	582
AAV-6	***************************************	581
22V-1	ACTTCCACCTCCATATTCTGGTGGAGACCACGGGGGTCAAATCCATGGTGCTGGGCCGCT	656
D D W - 2	A G. CG.G. C A C G	642
AAV-6		641
D D 77 7	TCCTGAGTCAGATTAGGGACAAGCTGGTGCAGACCATCTACCGCGGGATCGAGCCGACCC	716
DDV-2	C.C. A. A. A. T GA . T	702
AAV-6		701
		776
AAV-1	TGCCCAACTGGTTCGCGGTGACCAAGACGCGTAATGGCGCCGGAGGGGGAACAAGGTGG	762
AAV-2	A	761
AAV-b		

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FIG 1B

D D W - 2	TGGACGAGTGCTACATCCCCAACTACCTCCTGCCCAAGACTCAGCCCGAGCTGCAGTGGGTT.G.CA.CTC	836 822 821
A 237-2	P19/TATA P19 RNA™ CGTGGACTAACATGGAGGAGTATATAAGCGCCTGTTTGAACCTGGCCGAGCGCAAACGGCTACTCGTCAGTTCGGAGACGG.C	896 882 881
AAV-2	TCGTGGCGCAGCACCTGACCCACGTCAGCCAGGACCCAGGAGCAGAACAAGGAGAATCTGA .GTGGTCGGAA	956 942 941
AAV-2 AAV-6	Rep_52/40 ACCCCAATTCTGACGCGCCTGTCATCCGGTCAAAAACCTCCGCGCGCTACATGGAGCTGG .TTGGA.ATACAGA	1002
AAV-2	TCGGGTGGCTGGTGGACCGGGGCATCACCTCCGAGAAGCAGTGGATCCAGGAGGACCAGG	1062
2211-2	CCTCGTACATCTCCTTCAACGCCGCTTCCAACTCGCGGTCCCAGATCAAGGCCGCTCTGGATGCATCT	1122
2 – ענת ת	ACAATGCCGGCAAGATCATGGCGCTGACCAAATCCGCGCCCGACTACCTGGTAGGCCCCGG.ATAGCTAC	1196 1182 1181
AAV-2 AAV-6	CTCCGCCCGCGGACATTAAAACCAACCGCATCTACCGCATCCTGGAGCTGAACGGCTACG AGCGTG.ATCC.GTGTTAAATTAAG	1242
AAV-2 AAV-6	AACCTGCCTACGCCGGCTCCGTCTTTCTCGGCTGGGCCCAGAAAAGGTTCGGGAAGCGCA .TCCAATG.CTAACAA	1302
AAV-2	ACACCATCTGGCTGTTTGGGCCGGCCACCACGGGCAAGACCAACATCGCGGAAGCCATCG	1376 1362 1361
AAV-2	CCCACGCCGTGCCCTTCTACGGCTGCGTCAACTGGACCAATGAGAACTTTCCCTTCAATGA.T	1422
AAV-2	ATTGCGTCGACAAGATGGTGATCTGGTGGGAGGAGGGCAAGATGACGGCCAAGGTCGTGG .CT	1496 1482 1481
DDV-2	AGTCCGCCAAGGCCATTCTCGGCGGCAGCAAGGTGCGCGTGGACCAAAAGTGCAAGTCGTGAAA	1556 1542 1541
77V-2	CCGCCCAGATCGACCCCACCCCCGTGATCGTCACCTCCAACACCAACATGTGCGCCGTGA .GAGT	1602

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FIG 1C

	TTGACGGGAACAGCACCACCTTCGAGCACCAGCAGCCGTTGCAGGACCGGATGTTCAAAT	1676
AAV-1	TTGACGGGAACAGCACCACCTTCGAGCACCAGCAGCCGTTGCAGGACCCGTTTGAGGACCAGCAGCAGCAGCCGTTGCAGGACCCGTTTGAGGACCCGTTTGAGGACCCGTTTGAGGACCCGTTTGAGGACCCGTTTGAGGACCCGTTTGAGGACCCGTTTGAGGACCCGTTTGAGGACCCGTTTGAGGACCAGCAGCAGCAGCAGCAGCAGCAGCAGCAGCAGC	1662
AAV-2	TCAGA	1661
AAV-6	***************************************	
	TO THE TOTAL CONTROL OF THE TOTAL ASSOCIATION OF THE TOTAL OF THE TOTA	1740
AAV-1	TTGAACTCACCCGCCGTCTGGAGCATGACTTTGGCAAGGTGTGTGT	1722
AAV-2	T	1721
AAV-6	T	
		1796
AAV-1	AGTTCTTCCGCTGGGCGCAGGATCACGTGACCGAGGTGGCGCATGAGTTCTACGTCAGAA	1782
22V-6	.CTGAA	1,01
	P40/TATA	1856
n n 37 – 1	AGGGTGGAGCCAACAAAAGACCCGCCCCCGATGACGCGGGGAGCCAAGCGGGGAGCCCAAGCGGGAGGAGCCAAGCAAGAGACAAAAAGACCCGCCCCCGATGACGAGAGAAAAAGACCCAAGCGGGAGACCAAGCGGGAGACCAAGCGGGAGACCAAGCGGGAGCCCAAGCGGGAGACAAAAAGACCCGCCCCCGATGACGAGAGAAAAAGACCAAACAAA	1942
ב-מות	AGGGTGGAGCCAACAAAAGACCCGCCCCCGATGACGCGGATAAAAGCCAACAAAAAAAA	1012
DDV-6	G	1041
AAV 0		
	P40 RNA	1016
n n sz 1		1910
AAV-1	CCTGCCCCTCAGTCGCGGATCCATCGACGTCAGACGCGGAAGGAGGTCCGTCAGACGCGGAAGGACGCGGAAGGACGCGGAAGGACGCGGAAGGACGCGGAAGGACGCGGAAGGACGCGGAAGGAAGGACGCGGAAGGACGCGGAAGGACGCGGAAGGACGCGGAAGGACGCGGAAGGACGCGGAAGGAAGGACGCGGAAGGAAGGACGCGGAAGGAAGGACGCGGAAGGAAGGACGCGGAAGGAAGGACGCGGAAGGAAGGACGCGGAAGGAAGGACGCGGAAGGAAGGACGCGGAAGGAAGGACGCGGAAGGAAGGACGCGGAAGGAAGGACGCGGAAGGAAGGACGCGGAAGGAAGGACGCGGAAGGAAGGACGAC	1899
AAV-2	TGCGAGTC.G	1901
AAV-6		
	· ~	
	CCGACAGGTACCAAAACAAATGTTCTCGTCACGCGGGCATGCTTCAGATGCTGTTTCCCT	1976
AAV-1	CCGACAGGTACCAAAACAAATGTTCTCGTCACGCGGGCATGCTTCACATGGTCACAAAACAAATGTTCTCGTCACGCGGGCATGCTTCACATGGTCACAGTCACACAAAAAAAA	1959
AAV-2	.A	1961
AAV-6	***************************************	
	GCAAGACATGCGAGAGAATGAATCAGAATTTCAACATTTGCTTCACGCACG	2036
AAV-1	GCAAGACATGCGAGAATGAATGAATTAA	2019
AAV-2	GCAAGACATGCGAGAGAATGAATCAGAATTTCAACATTTGCTTCAGCATCAGCAA.CATCTACAA	2021
AAV-6		
	TO A PROMOTE A P	2093
AAV-1	ACTGTTCAGAGTGCTTCCCCGGCGTGTCAGAATCTCAACCCCTTCT GTC A.A.G	2076
AAV-2	ACTGTTCAGAGTGCTTCCCCGGCGTGTCAGAATCTCAACCGGTCTTCTGTCA.A.G	2078
AAV-6	AT	
	TOTAL TOTAL CONTROL OF THE STATE OF THE STAT	2153
AAV-1	CGTATCGGAAACTCTGTGCCATTCATCATCTGTGGGCGGGC	2133
AAV-2	AGCTAA.CAAAAG	2138
AAV-6	AGCTAA.CAAAAA	
	Ren 78 stop	
	TO A CONTROL OF THE PROPERTY O	2213
AAV-1	CGGCCTGCGATCTGGTCAACGTGGACCTGGATGACTGTGTTTCTGAGCTGTGATGACTGTGTTTCTGAGCTGAGCTGGACTGGATGACTGTGTTTCTGAGCTGGACTGTGTTTCTGAGCTGGACTGGATGACTGTGTTTCTGAGCTGGACTGTGTTTCTGAGCTGGACTGGATGACTGTGTTTCTGAGCTGGACTGGATGACTGTGTTTCTGAGCTGAGCTGGACTGGACTGGACTGGACTGGACTGGACTGGACTGGACTGGACTGGACTGGACTGGACTGGACTGGACTGGACTGGACTGGACTGGACTGGACTGACT	2193
AAV-2	.TTTTTCA.C.I	2193
AAV-6	5TTTT	
	Rep. 68 5	stop
	V VP1 AAACCAGGTATGGCTGCCGATGGTTATCTTCCAGATTGGCTCGAGGACAACCTCTCTGAC CT	2273
AAV-	AAACCAGGTATGGCTGCCGATGGTTATCTTCCAGATTGGCTCGAGGACAACTCT	2253
AAV-	AAACCAGGTATGGCTGCCGATGGTTATCTTCCAGATTGGGTGATGGTTATCTTCCAGATTGGTTGCTCAGATTGGTTATCTTCCAGATTGGTTATCTTTCCAGATTGGTTATCTTTCCAGATTGGTTATCTTTCCAGATTGGTTATCTTTCCAGATTGGTTATCTTTCCAGATTGGTTATCTTTCCAGATTGGTTATCTTTCCAGATTGGTTATCTTTCCAGATTGGTTATCTTTCCAGATTGGTTATCTTTCCAGATTGGTTATCTTTCCAGATTGGTTATCTTCCAGATTGGTTATCTTTCCAGATTGGTTATCTTTCCAGATTGGTTATCTTTCCAGATTGGTTATCTTTCCAGATTGGTTATCTTTCCAGATTGGTTATCTTTCCAGATTGGTTATCTTTCCAGATTGGTTATCTTTCCAGATTGGTTATCTTTCCAGATTGGTTATCTTTCCAGATTGGTTATCTTTCCAGATTGGTTATCTTTCCAGATTGGTTATCTTTCCAGATTATCTTTCAGATTATCTTTCAGATTATCTTTCAGATTATCTTTCAGATTATCTTTCAGATTATCTTTCAGATTATCTTTTCAGATTATCTTTCAGATTATCTTTCAGATTATCTTTCAGATTATCTTTTCAGATTATCTTTTTTTT	225B
AAV-	2TAC	
		A 2333
AAV-	GGCATTCGCGAGTGGTGGGACTTGAAACCTGGAGCCCGAAGCCCAAAGCCAACCAGCA	3 2313
AAV-	GGCATTCGCGAGTGGTGGGACTTGAAACCTGGAGCCCCGAAGCCAAACCAAACCAAAACAAAAAAAA	2318
AAV-	2AAA.AC	
-Vaa	1 AAGCAGGACGACGGCCGGGGTCTGGTGCTTCCTGGCTACAAGTACCTCGGACCCTTCAA	2377
AAV-	1 AAGCAGGACGACGGCGGGGTCTGGTGCTTCCTGGCTACAAGTACGTCCGCCACAAGTACGTCCAAGTACAAGTACGTCCAAGTACAAGTACGTCCAAGTACAAGTACGTCCAAGTACAAGTACGTCCAAGTACAAGTACGTCCAAGTACAAGTACGTCCAAGTACAAGTACGTCCAAGTACAAGTACGTCCAAGTACAAGTACGTCCAAGTACAAGTACGTCCAAGTACAAGTACGTCCAAGTACAAGTACGTCCAAGTACAAGTACGTCCAAGTACAAGTACGTCCAAGTACAAGTACGTCCAAGTACAAGTACAAGTACAAGTACAAGTACAAGTACAAGTACAAGTACAAGTACAAGTACAAGTACAAGTACAAGTACAAAGTACAAGTACAAAGTACAAGTACAAGTACAAAGTACAAAGTACAAGTACAAAGTACAAAGTACAAAGTACAAAGTACAAAGTACAAAGTACAAAGTACAAAGTACAAAGTACAAAAAAAA	. 23/3
DAY-	2 C.TA	. 23/
WW A	V 1111111111111111	

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FIG 1D

AAV-1	GGACTCGACAAGGGGGACCCGTCAACGCGGCGGACGCAGCGGCCCTCGAGCACGACAAG	2453
DAV-2	AGAACA	2433
777-6	T	2438
rusv u		
B B 17 1	GCCTACGACCAGCAGCTCAAAGCGGGTGACAATCCGTACCTGCGGTATAACCACGCCGAC	2513
MAV-1	GG.CAGC.ACCAAC	2493
AAV-2	A.AGCGTTGCGT	2408
AAV-6	A.AGCGTTGCGT	2430
		2572
AAV-1	${\tt GCCGAGTTTCAGGAGCGTCTGCAAGAAGATACGTCTTTTGGGGGCAACCTCGGGCGAGCA}$	23/3
AAV-2	GA	2555
AAV-6	CTGC	2558
AAV-1	GTCTTCCAGGCCAAGAAGCGGGTTCTCGAACCTCTCGGTCTGGTTGAGGAAGGCGCTAAG	2633
2 AV-2	G. A. A	2613
AAV-6	T.TT	2618
	VP2	
n n t/_ 1	ACGCTCCTGGAAAGAAACGTCCGGTAGAGCAGTCGCCACAAGAGCCAGACTCCTCCTCG	2693
MAN - I	GAGA.GCTTGTG	2673
AAV-Z	TGAC.TGGACAA	2678
AAV-6	TGAC.I	20.0
	GGCATCGGCAAGACAGGCCAGCCCGCTAAAAAGAGACTCAATTTTGGTCAGACTGGC	2753
AAV-1	.A.CAG.GA	2733
AAV-2	A.CAG.G	2720
AAV-6	T	2130
	GACTCAGAGTCAGTCCCCGATCCACAACCTCTCGGAGAACCTCCAGCAACCCCCGCTGCT	2017
AAV-1	GACTCAGAGTCAGTCCCCGATCCACAACCTCTCGGAGAACCTCCAGCAACCTCTCGGAGAACCTCTCAGCAAACCTCTCGGAGAACCTCTCAGCAAACCTCTCGGAGAACCTCTCAGCAAACCTCAGCAAACCTCTCAGCAAACCTCAGCAAACCTCAAACCTCAGCAAACCTCAGCAAACCTCAGCAAACCTCAGCAAACCTCAGCAAACCTCAGCAAACCTCAGCAAACCTCAGCAAACCTCAGCAAACCTCAGCAAACCTACAAACCTACAAACCTACAAACCTACAAACCTACAAACCTACAAACCTACAAACCTACAAACCTACAAACCTACAAACCTACAAACCTACAAAACCTACAAAACCTACAAAACCTACAAAACCTACAAAACAAACCTACAAAACAAACCTACAAAACAAAACAAAACAAAACAAAAAA	2703
AAV-2	GCATCCGC.GAGTG.	2/93
AAV-6	TGCCCAAG.ATAG	2/98
	VP3	
AAV-1	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGCACCAATGGCAGACAATAACGAAGGC	2873
2-V44	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGCACCAATGGCAGACAATAACGAAGGC	2853
2-V44	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGCACCAATGGCAGACAATAACGAAGGC	2853
AAV-2 AAV-6	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGCACCAATGGCAGACAATAACGAAGGC CAAGAAGG.	2853 2858
AAV-2 AAV-6	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGCACCAATGGCAGACAATAACGAAGGC CAAGAAGAGG.	2853 2858 2933
AAV-2 AAV-6	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGCACCAATGGCAGACAATAACGAAGGC CAAGAAGAGG.	2853 2858 2933
AAV-2 AAV-6 AAV-1 AAV-2	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGCACCAATGGCAGACAATAACGAAGGC CAAGAAGAGG.	2853 2858 2933 2913
AAV-2 AAV-6 AAV-1 AAV-2	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGCACCAATGGCAGACAATAACGAAGGC CAAGAAGAGG.	2853 2858 2933 2913
AAV-2 AAV-6 AAV-1 AAV-2 AAV-6	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGCACCAATGGCAGACAATAACGAAGGC C. A. A. G. A. A. G. G. C. A. A. G. G. C. A. A. G. C. A. A. G. G. C. G.	2853 2858 2933 2913 2918
AAV-2 AAV-6 AAV-1 AAV-2 AAV-6	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGCACCAATGGCAGACAATAACGAAGGC C. A. A. G. A. A. A	2853 2858 2933 2913 2918 2993
AAV-2 AAV-6 AAV-1 AAV-6 AAV-1	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGACCAATGGCAGACAATAACGAAGGC CAAGAAGAAGGAA	2853 2858 2933 2913 2918 2993 2973
AAV-2 AAV-6 AAV-1 AAV-6 AAV-1	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGCACCAATGGCAGACAATAACGAAGGC C. A. A. G. A. A. A	2853 2858 2933 2913 2918 2993 2973
AAV-2 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGCACCAATGGCAGACAATAACGAAGGC C. A. A. G. A. A. A. G. G.	2853 2858 2933 2913 2918 2993 2973 2978
AAV-2 AAV-6 AAV-1 AAV-6 AAV-1 AAV-2 AAV-6	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGCACCAATGGCAGACAATAACGAAGGC C. A. A. G. A. A. A	2853 2858 2933 2913 2918 2993 2973 2978 3053
AAV-2 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-6 AAV-1	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGCACCAATGGCAGACAATAACGAAGGC C. A. A. G. A. A. A	2853 2858 2933 2913 2918 2993 2973 2978 3053 3030
AAV-2 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-6 AAV-1	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGCACCAATGGCAGACAATAACGAAGGC C. A. A. G. A. A. A	2853 2858 2933 2913 2918 2993 2973 2978 3053 3030
AAV-1 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGCACCAATGGCAGACAATAACGAAGGC C. A. A. G. A. A. A. G. G. A. A. G. G. G. G. G. A. A. G. A. A. G. G. A. A. G.	2853 2858 2933 2913 2918 2993 2973 2978 3053 3030 3038
AAV-2 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGACCAATGGCAGACAATAACGAAGGC CAAGAAA	2853 2858 2933 2913 2918 2973 2978 3053 3030 3038 3113
AAV-1 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGACCAATGGCAGACAATAACGAAGGC C. A. A. G. A. A. A. A. G. G. A. A. G. G. A. A. G. A. A. A. G. G.	2853 2858 2933 2913 2918 2973 2978 3053 3030 3038 3113 3090
AAV-1 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGACCAATGGCAGACAATAACGAAGGC CAAGAAA	2853 2858 2933 2913 2918 2973 2978 3053 3030 3038 3113 3090
AAV-2 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGACCAATGGCAGACAATAACGAAGGC CAAGAAA	2853 2858 2933 2913 2918 2993 2973 2978 3053 3030 3038 3113 3098
AAV-1 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGCACCAATGGCAGACAATAACGAAGGC C. A. A. G. A. A. A	2853 2858 2933 2913 2918 2993 2973 2978 3053 3030 3038 3113 3090 3098
AAV-1 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGCACCAATGGCAGACAATAACGAAGGC C. A. A. G. A. A. A	2853 2858 2933 2913 2918 2993 2978 3053 3030 3038 3113 3090 3098 3173 3150
AAV-1 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGCACCAATGGCAGACAATAACGAAGGC C. A. A. G. A. A. A	2853 2858 2933 2913 2918 2993 2978 3053 3030 3038 3113 3090 3098 3173 3150
AAV-2 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGACCAATGGCAGACAATAACGAAGGC CAAGAAA	2853 2858 2933 2913 2918 2973 2978 3053 3030 3038 3113 3090 3098 3158
AAV-1 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGCACCAATGGCAGACAATAACGAAGGC C. A. A. G. A. A. A. A	2853 2858 2933 2913 2918 2993 2973 2978 3053 3030 3038 3113 3090 3173 3150 3158
AAV-1 AAV-1 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGCACCAATGGCAGACAATAACGAAGGC C. A. A. G. A. A. A. A	2853 2858 2933 2913 2918 2993 2973 2978 3053 3030 3038 3113 3090 3158 3233 3233
AAV-1 AAV-1 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1 AAV-2 AAV-6 AAV-1	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGACCAATGGCAGACAATAACGAAGGC CAAGAAA	2853 2858 2933 2913 2918 2993 2973 2978 3053 3030 3038 3113 3090 3158 3233 3233

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FIG 1E

スカリーフ	ACCAGCACGGTTCAAGTCTTCTCGGACTCGGAGTACCAGCTTCCGTACGTCCTCGGCTCT	3270
2-1/4	GCGCACCAGGGCTGCCTCCCTCCGTTCCCGGCGGACGTGTTCATGATTCCGCAATACGGCT.A.AGA.ACG.G.AGTAGG.G.G.G.G.G.G.G.G.G.G.G.G	3330
DDV-2	TACCTGACGCTCAACAATGGCAGCCAAGCCGTGGGACGTTCATCCTTTTACTGCCTGGAACCGCGTGAACTA	3390
AAV-2 AAV-6	TATTTCCCTTCTCAGATGCTGAGAACGGGCAACAACTTTACCTTCAGCTACACCTTTGAGCTTTT	3450 3458
2 - W44	GAAGTGCCTTTCCACAGCAGCTACGCGCACAGCCAGAGCCTGGACCGGCTGATGAATCCTCTTCTC	3510
2 2 37 - 2	CTCATCGACCAATACCTGTATTACCTGAACAGAACTCAAAATCAGTCCGGAAGTGCCCAA	3570
AAV-2	AACAAGGACTTGCTGTTTAGCCGTGGGTCTCCAGCTGGCATGTCTGTTCAGCCCAAAAACCC.GTCAAGGC.T.ATCT.AG.CCGGAG.GAGATCGG.ACT.T.GG	3630
AAV-2	TGGCTACCTGGACCCTGTTATCGGCAGCAGCGGGTTTCTAAAACAAAAACAGACAACAACTCCA.A.CAGTCTG.GT	3690
AAV-2	AACAGCAATTTTACCTGGACTGGTGCTTCAAAATATAACCTCAATGGGCGTGAATCCATCTG.A.ACT.GAA.CGCCCA.ACTC.GCTA	3750
AAV-2	ATCAACCCTGGCACTGCTATGGCCTCACACAAAGACGACGAAGACAAGTTCTTTCCCATG G.GTGGC.CCAAGCGTAT.CAA	3810
AAV~2	AGCGGTGTCATGATTTTTGGAAAAGAGAGCGCCGGAGCTTCAAACACTGCATTGGACAATGTC.CGGC.AGT.A.AGAAAATGTGAACA.TAGGG	3893 3870 3878
AAV-2	GTCATGATTACAGACGAAGAGGAAATTAAAGCCACTAACCCTGTGGCCACCGAAAGATTTCGG.A.ACTCTGGCAG.A.	3930
AAV-2	GGGACCGTGGCAGTCAATTTCCAGAGCAGCAGCACCAGACCCTGCGACCGGAGATGTGCATTT.T.AT.TACCCAGAG.C.AG.ATCCA.CTC	3990
AAV-2	GCTATGGGAGCATTACCTGGCATGGTGTGGCAAGATAGAGACGTGTACCTGCAGGGTCCC A.ACAAC.TTC.TACGCTT.	4050

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FIG 1F

A A SE 1	ATTTGGGCCAAAATTCCTCACACAGATGGACACTTTCACCCGTCTCCTCTTATGGGCGGC	4133
WWA-1	CAGAGCT	4110
AAV-2		4118
AAV-6		
3 3 W 1	TTTGGACTCAAGAACCCGCCTCCTCAGATCCTCATCAAAAACACGCCTGTTCCTGCGAAT	4193
MMV-1		4170
AAV-2	TC	4178
AAV-6		
ΛΑ W_1	CCTCCGGCGGAGTTTTCAGCTACAAAGTTTGCTTCATTCA	4253
77V-2	TA.CACCCAGTGGCAGG	4230
ARV-E	AG	4238
AAV-0	A	
AAV-1	CA-AGTGAGTGTGGAAATTGAATGGGAGCTGCAGAAAGAAAACAGCAAGCGCTGGAATCC	4312
AAV-2	cgcc	4290
DDV-6	A	4297
AAV-1	CGAAGTGCAGTACACATCCAATTATGCAAAATCTGCCAA-CGTTGATTTTACTGTGGACA	4371
AAV-2	A.TTCCAACGTTTGCT.	4350
AAV-6	TTC	4356
AAV-1	${\tt ACAATGGACTTATACTGAGCCTCGCCCCATTGGCACCCGTTACCTTACCCGTCCCCTG\overline{T}}$	4431
DAV-2	CTCG.GT.AA.AGTAAT	4410
AAV-6		4416
	VP1-3 stop PolyA signal	
AAV-1	AATTACGTGTTAATCAATAAACCGGTTGATTCGTTTCAGTTGAACTTTGGTCTCCTGTCC	4491
D D W-2	G T T. A	4470
AAV-6	GTAGAG	4476
AAV-1	TTCTTATCTTATC-GGTTACCATGGTTAT-AGCTTACACATTAACTGCTTGGTTGCGC	4547
AAV-2	TC.TTATCCGTAGAAGT.GC.TGG.G.GAA.CATTA	4530
AAV-6	ATCA.CA.C-C.GAA	4533
		4 6 8 8
AAV-1	${\tt TTCGCGATAAAAGACTTACGTCATCGGGttacccctagtgatggagttgcccactccctc}$	4607
AAV-2	ACTA.A.gg.a	4570
AAV-6	at	4572
AAV-1	${\tt tctgcgcgctcgctcggtcggtggggccggcagagcaga$	4667
AAV-2	.cacacagccaggcagc.c.gg,	4630
AAV-6	.aggg	4632
AAV-1	tggtccgcaggccccaccgagcgagcgagcgcagagagggagtgggcaa	4718
AAV-2	cc.g.gctgt	4681
AAV-6		4683

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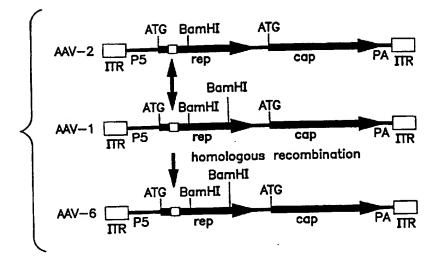
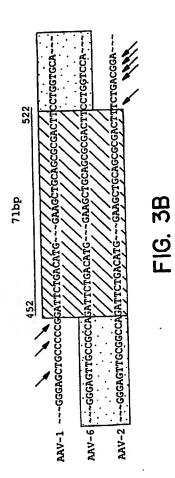
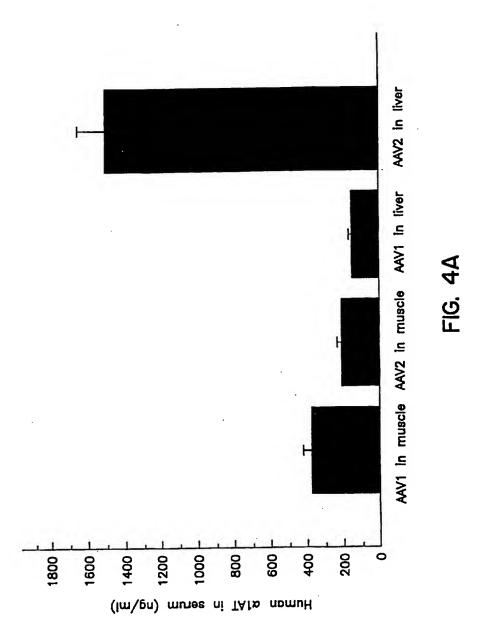


FIG. 3A

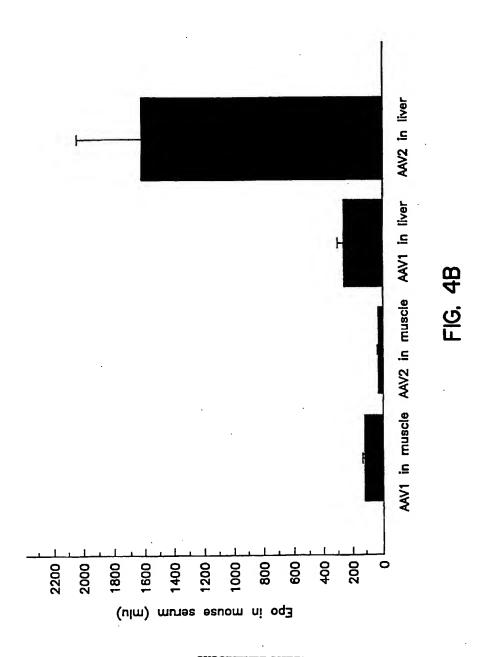


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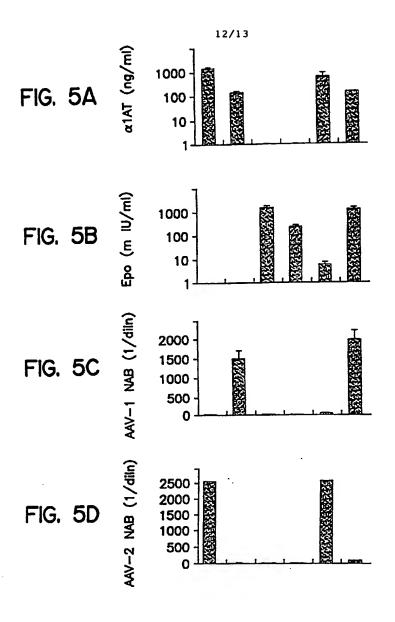


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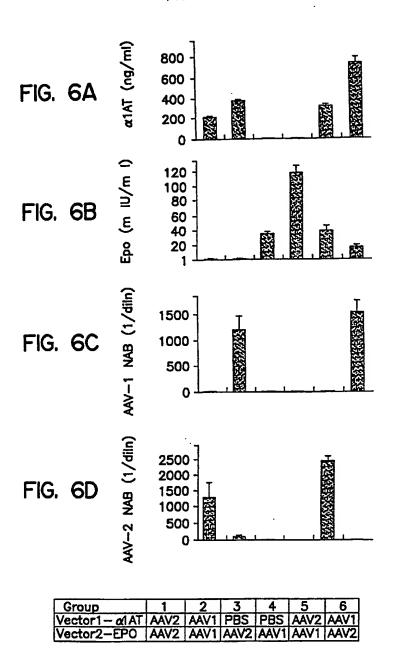
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Vector2-EPO	AAV2	AAV1	AAV2	AAV1	AAV1	AAV2

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SEQUENCE LISTING

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<110> Wilson, James M. Xiao, Weidong The Trustees of the University of Pennsylvania <120> Adeno-Associated Virus Serotype I Nucleic Acid Sequences, Vectors and Host Cells Containing Same <130> GNVPN.031PCT <140> <141> <150> 60/107,114 <151> 1998-11-05 <160> 20 <170> PatentIn Ver. 2.0 <210> 1 <211> 4718 <212> DNA <213> AAV-1 <220> <221> CDS <222> (335)..(2206) <220> <221> CDS <222> (2223)..(4430) <400> 1 ttgcccactc cctctctgcg cgctcgctcg ctcggtgggg cctgcggacc aaaggtccgc 60 agacggcaga gctctgctct gccggcccca ccgagcgagc gagcgcgcag agagggagtg 120 ggcaactcca tcactagggg taatcgcgaa gcgcctccca cgctgccgcg tcagcgctga 180 cgtaaattac gtcatagggg agtggtcctg tattagctgt cacgtgagtg cttttgcgac 240 attttgcgac accacgtggc catttagggt atatatggcc gagtgagcga gcaggatctc 300 cattttgacc gcgaaatttg aacgagcagc agcc atg ccg ggc ttc tac gag atc 355

1

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Met Pro Gly Phe Tyr Glu Ile

gtg	atc	aag	gtg	ccg	agc	gac	ctg	gac	gag	cac	ctg	ccg	ggc	att	tct	403
Val	Ile	Lys	Val	Pro	Ser	Asp		Asp	Glu	His	Leu		Gly	Ile	Ser	
		10					15					20				
σac	tca	ttt	ata	agc	tgg	gtg	gcc	gag	aag	gaa	tgg	gag	ctg	ccc	ccg	451
Asp	Ser	Phe	Val	Ser	Trp	Val	Ala	Glu	Lys	Glu	Trp	Glu	Leu	Pro	Pro	
•	25					30					35					
nat	tct	σac	atσ	σat	cta	aat	ctg	att	gag	cag	gca	ccc	ctg	acc	gtg	499
Asp	Ser	Asp	Met	Asp	Leu	Asn	Leu	Ile	Glu	Gln	Ala	Pro	Leu	Thr	Val	
40		•		•	45					50					55	
														~+~	nat.	547
gcc	gag	aag	ctg	cag	cgc	gac	Dha	ctg	gcc	Cla	Trn	ara ara	Dra.	gtg Val	Ser	34,
Ala	GLU	Lys	ren	60	Arg	Asp	PHE	Ten	65	GIII	itp	ALG	Ary	70	501	
				00					0.5					. •		
aag	gcc	ccg	gag	gcc	ctc	ttc	ttt	gtt	cag	ttc	gag	aag	ggc	gag	tcc	595
Lys	Ala	Pro	Glu	Ala	Leu	Phe	Phe	Val	Gln	Phe	Glu	Lys	Gly	Glu	Ser	
			75					80					85			
tac	ttc	cac	ctc	cat	att	cta	ata	gag	acc	acq	aga	qtc	aaa	tcc	atg	643
														ser		
-,-	2	90					95				•	100	_			
gtg	ctg	ggc	cgc	ttc	ctg	agt	cag	att	agg	gac	aag	ctg	gtg	cag	acc	691
Val	Leu	Gly	Arg	Phe	Leu	Ser	Gln	Ile	Arg	Asp		Leu	Val	Gln	Thr	
	105					110					115					
atc	tac	cac	aaa	atc	gag	ccq	acc	ctg	ccc	aac	tgg	ttc	gcg	gtg	acc	739
Ile	Tyr	Arg	Gly	Ile	Glu	Pro	Thr	Leu	Pro	Asn	Trp	Phe	Ala	Val	Thr	
120	_	-			125					130					135	
																787
aag	acg	cgt	aat	ggc	gcc	gga	ggg	999	aac	aag	gra	g cg	Den	gag Glu	Cve	, , ,
Lys	Thr	Arg	Asn	140	Ald	GIY	GIY	Gry	145	цуз	Val	var	Д	150	.,.	
				140												
tac	atc	ccc	aac	tac	ctc	ctg	ccc	aag	act	cag	ccc	gag	ctg	cag	tgg	835
														Gln		
•			155					160					165			
gcg	tgg	act	aac	atg	gag	gag	tat	ata	agc	gcc	tgt	ttg	aac	ctg	gcc	883
Ala	Trp		Asn	Met	Glu	Glu		Ile	Ser	Ala	Cys	Leu 180	AST	Leu	ALA	
		170					175					190				
gag	cac	aaa	caa	ctc	gta	gcg	cag	cac	ctg	acc	cac	gtc	agc	cag	acc	931
Glu	Arg	Lys	Arg	Leu	Val	Ala	Gln	His	Leu	Thr	His	Val	Ser	Gln	Thr	
	185	-				190					195					

cag	gag	cag	aac	aag	gag	aat	ctg	aac	ccc	aat	tct	gac	geg	cct	gtc	979
Gln	Glu	Gln	Asn	Lys	Glu	Asn	Leu	Asn	Pro	Asn	Ser	Asp	Ala	Pro	Val	
200				_	205	1				210)	_			215	
atc	caa	tca	aaa	acc	tcc	aca	cac	tac	ato	σασ	cta	ato	aac	tac	ctg	1027
							_		_		_	-			Leu	
	,		-,-	220				-,-	225				,	230		
ata	dac	caa	aac	arc	acc	tee	дад	аас	cad	taa	ato	Cad	gag	gac	cag	1075
	-							_	-			-		_	Gln	
		,	235					240					245	_		
acc	tca	tac	atc	tcc	ttc	aac	acc	act	tcc	aac	tca	caa	tcc	cad	atc	1123
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		230					200									
aan	qcc	act	cta	gac	aat	acc	aac	220	atc	ato	aca	cta	acc	222	tcc	1171
-	Ala	-	_	-		-		_		-		•				
LyJ	265					270	01,	_,_			275	200	••••	٠,٠	501	
	203					270					213					
aca	ccc	a = c	tac	cta	at a	aac	ccc	act	cca		aca	~~~	att	222	366	1219
-	Pro	-		-	-			-	-			_				1213
280	FIO	нар	TYL	rea	285	GIY	FIO	MIG	FIU	290		Asp	116	Lys	295	
200					203					250					293	
	cgc	ato	tac	cac	ato	cra	a a a	cta	220	990	rac	722	cct	~~~	tac	1267
	Arg			-		-		_				-		-		1207
	y	110	- 3 -	300			-		305	- -,	. , .	OIU	110	310	- y -	
				500					505					310		
acc	ggc	tee	a+c		crc	000	taa	~~~	c a 4	222	200	++~		225	686	1315
-	Gly		-						-					_	-	1313
мта	Gry	261	315	FILE	rea	Gry	TTP	320	GIH	Lys	ALY	FILE	325	Lys	ALG	
			212					320					323			
	acc					~~~		~~~	200	200	~~~		200		250	1363
				-			-	-		-	• •	_				1363
ASII	Thr	330	пр	Leu	Pne	GLY	335	MIG	Int	THE	GIY	-	THE	ASD	116	
		330					333					340				
aca		~~~	200	~~~	~~~		ata		++-		~~~		a+ a		+~~	1411
	gaa Glu	-		-		-						-	-			1411
мта	345	мта	116	жта	NIS	350	Vai	PIO	rne	ıyı	355	cys	Val	ASII	1LP	
	343					330					333					
	+	40 5	225				+	+		~+~	~~~					1450
	aat							-	_	-	_	-	-			1459
	Asn	GIU	ASN	rne		rne	Asn	Asp	cys		Asp	ràz	met	vai		
360					365					370					375	
																1507
	tgg															1507
rrp	Trp	GIU		GIA	гÀг	met	Thr		Lys	val	val	Glu	ser	Ala	гÀг	

															g tcg	1555
Ala	110	e Le			y Se	Ly	s Val			Asp	Gli	n Lys			s Ser	
			39	5				400)				40	5		
tce		cad	r atr	na/		. acc		· ata	, atc	ate		. +			c aac	1603
				-						_					r Asn	1003
		410					415					420				
atç	tgo	gco	gto	att	gac	ggç	aac	ago	acc	acc	tto	gag	cad	ca	g cag	1651
Met	Cys	Ala	val	Ile	Asp	Gly	Asn	Ser	Thr	Thr	Phe	e Glu	His	5 G1:	n Gln	
	425	j.				430	t				435	5				
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GIU	pro 505	Lys	Arg	Ala	Cys	Pro 510	Ser	Val	Ala	Asp		Ser	Thr	Ser	Asp	
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qcq	gaa	gga	act	cca	ata	gac	ttt	acc	gac	agg	tac	caa	aac	aaa	tat	1939
									-			Gln			-	1,0,
520					525				•	530	-			•	535	
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Ser	Arg	His	Ala		Met	Leu	Gln	Met		Phe	Pro	Cys	Lys		Cys	
				540					545					550		
gag	aga	ata	aat	caa	aat	ttc	aac	att	tac	tte	aca	cac	aaa	acc	aga	2035
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	_		555					560	•				565			
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Asp	Cys		Glu	Cys	Phe		-	Val	Ser	Glu	Ser	Gln	Pro	Val	Val	
		570					575					580				

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	Ala				-	Cys	-	-	-	-	Leu	-			gac Asp 615	2179
-	-	gac Asp	_	-	Ser				atg	actt	aaa	ccag	M		ct gcc la Ala	
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-				-	-		Pro		-	-	_			_	aac Asn	2327
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		gga Gly													gcg Ala	2423
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		ggt Gly 710										-		-	gag Glu	2519
	-	gag Glu	-	-											ggg Gly	2567
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		o Al					Asn					Gl	-	c tca p Ser	
	va:					Pro			_		Pro	_		ccc r Pro	2807
Ala					Thr					Gly				a atg Met 835	2855
				Gly					Gly		-			a aat / Asn)	2903
			Ser										Thr	agc Ser	2951
		Trp											-	caa Gln	2999
	Ser													ggc Gly	3047
			tgg Trp							-			-		3095
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			ctc Leu												3191
			gat Asp		Val					Asn					3239

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	y Cys Leu Pr		g gcg gac gtg o Ala Asp Val	
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		Glu Tyr Phe	cct tct cag Pro Ser Gln 1025	•
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-	Tyr Leu Tyr	-	aga act caa Arg Thr Gln	
	Asn Lys Asp		agc cgt ggg Ser Arg Gly 1090	
			cct gga ccc Pro Gly Pro 1105	-
Gln Arg Val		Lys Thr Asp	aac aac aac Asn Asn Asn 1120	
Trp Thr Gly			aat ggg cgt Asn Gly Arg	-
			aaa gac gac (Lys Asp Asp (

gac	aag	ttc	ttt	ccc	atg	agc	ggt	gtc	atg	att	ttt	gga	aaa	gag	, agc	3863
Asp	Lys	Phe				Ser	Gly			Ile	Phe	Gly	Lys		Ser	
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-		-				-								-	Glu	
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	-														acc	3959
31u		11e 1190	_	WIG	THE		PF0 1195		AIa	mı		1200		GIY	Thr	
gtg	gca	gtc	aat	ttc	cag	agc	agc	agc	aca	gac	cct	gcg	acc	gga	gat	4007
/al	Ala	Val	Asn	Phe			Ser	Ser	Thr	-			Thr	Gly	Asp	
	1205				:	1210					1215					
ata	cat	act	ato	gga	gca	tta	cct	aac	ato	ata	taa	caa	gat	aσa	gac	4055
-															Asp	
122	0				1225				:	1230					1235	
		-	-					-						-	gga	4103
/al	Tyr	Leu		-	Pro	Ile	Trp		-	Ile	Pro	His		-	Gly	
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cac	ttt	cac	ccg	tct	cct	ctt	atg	ggc	ggc	ttt	gga	ctc	aag	aac	ccg	4151
iis	Phe	His	Pro	Ser	Pro	Leu	Met	Gly	Gly	Phe	Gly	Leu	Lys	Asn	Pro	
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		_	atc Ile													4199
		1270					275					1280				
_			tca	-		_										4247
		Phe	Ser	Ala		-	Phe	Ala	Ser			Thr	Gln	Tyr	Ser	
	1285				1	290					1295					
ca	gga	caa	gtg	agt	gtg	gaa	att	gaa	tgg	gag	ctg	cag	aaa	gaa	aac	4295
			Val								-	_		-		
306)			1	305				3	310				:	1315	
			tgg -											-		4343
er	Lys	Arg	Trp	Asn 320	Pro	GLu	۷al		Tyr 325	Thr	ser	Asn	-	Ala 1330	Lys	
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ct	gcc	aac	gtt	gat	ttt	act	gtg	gac	aac	aat	gga	ctt	tat	act	gag	4391
	-		Val	-												
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Thr	Gln	Pro	Glu	Leu 165		Trp	Ala	Trp	Thr 170		Met	Glu	Glu	Tyr 175	Ile
Ser	Ala	Cys	Leu 180		Leu	Ala	Glu	Arg 185	Lys	Arg	Leu	Val	Ala 190		His
Leu	Thr	His 195	Val	Ser	Gln	Thr	Gln 200	Glu	Gln	Asn	Lys	G1u 205		Leu	Asn
Pro	Asn 210		Asp	Ala	Pro	Val 215		Arg	Ser	Lys	Thr 220		Ala	Arg	Tyr
Met 225	Glu	Leu	Val	Gly	Trp 230	Leu	Val	Asp	Arg	Gly 235	Ile	Thr	Ser	Glu	Lys 240
Gln	Trp	Ile	Gln	Glu 245	Asp	G1r.	Ala	Ser	Tyr 250		Ser	Phe	Asn	Ala 255	Ala
Ser	Asn	Ser	Arg 260	Ser	Gln	Ile	Lys	Ala 265	Ala	Leu	Asp	Asn	Ala 270	Gly	Lys
Ile	Met	Ala 275	Leu	Thr	Lys	Ser	Ala 280	Pro	Asp	Tyr	Leu	Val 285	Gly	Pro	Ala
Pro	Pro 290	Ala	Asp	Ile	Lys	Thr 295	Asn	Arg	Ile	туг	Arg 300	Ile	Leu	Glu	Leu
Asn 305	Gly	Tyr	Glu	Pro	Ala 310	туг	Ala	Gly	Ser	Val 315	Phe	Leu	G1 y	Trp	Ala 320
Gln	Lys	Arg	Phe	Gly 325	Lys	Arg	Asn	Thr	11e 330	Trp	Leu	Phe	Gly	Pro 335	Ala
Thr	Thr	Gly	Lys 340	Thr	Asn	Ile	Ala	Glu 345	Ala	Ile	Ala	His	Ala 350	Val	Pro
Phe	Tyr	Gly 355	Cys	Val	Asn	Trp	Thr 360	Asn	Glu	Asn	Phe	Pro 365	Phe	Asn	Asp
Cys	Val 370	Asp	Lys	Met		Ile 375	Trp	Trp	Glu		Gly 380	Lys	Met	Thr	Ala
Lys 385	Val	Val	G1 u	Ser	Ala 390	Lys	Ala	lle	Leu	Gly 395	Gly	Ser	Lys	Val	Arg 400

Val Asp Gln Lys Cys Lys Ser Ser Ala Gln Ile Asp Pro Thr Pro Val 405 410 415

- Ile Val Thr Ser Asn Thr Asn Met Cys Ala Val Ile Asp Gly Asn Ser 420 425 430
- Thr Thr Phe Glu His Gln Gln Pro Leu Gln Asp Arg Met Phe Lys Phe 435 440 445
- Glu Leu Thr Arg Arg Leu Glu His Asp Phe Gly Lys Val Thr Lys Gln 450 455 460
- Glu Val Lys Glu Phe Phe Arg Trp Ala Gln Asp His Val Thr Glu Val 465 470 475 480
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- Pro Asp Asp Ala Asp Lys Ser Glu Pro Lys Arg Ala Cys Pro Ser Val 500 505 510
- Ala Asp Pro Ser Thr Ser Asp Ala Glu Gly Ala Pro Val Asp Phe Ala 515 520 525
- Asp Arg Tyr Gln Asn Lys Cys Ser Arg His Ala Gly Met Leu Gln Met 530 535 540
- Leu Phe Pro Cys Lys Thr Cys Glu Arg Met Asn Gln Asn Phe Asn Ile 545 550 555 560
- Cys Phe Thr His Gly Thr Arg Asp Cys Ser Glu Cys Phe Pro Gly Val
- Ser Glu Ser Gln Pro Val Vai Arg Lys Arg Thr Tyr Arg Lys Leu Cys 580 585 590
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Lys	Ala	Asn 35		Gln	Lys	Gln	Asp 40		Gly	Arg	Gly	Leu 45		Leu	Pro
Gly	Туг 50	-	Туr	Leu	Gly	Pro 55		Asn	Gly	Leu	Asp 60	-	Gly	Glu	Pro
Val 65	Asn	Ala	Ala	Asp	Ala 70	Ala	Ala	Leu	Glu	His 75	Asp	Lys	Ala	Tyr	Asp 80
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Asp	Ala	Glu	Phe		Glu	Arg	Leu	Gln 105	Glu	Asp	Thr	Ser	Phe 110	Gly	Gly
Asn	Leu	Gly 115	Arg	Ala	Val	Phe	Gln 120	Ala	Lys	Lys	Arg	Val 125	Leu	Glu	Pro
Leu	Gly 130	Leu	Val	Glu	Glu	Gly 135	Ala	Lys	Thr	Ala	Pro 140	Gly	Lys	Lys	Arg
Pro 145	Val	Glu	Gln	Ser	Pro 150	Gln	Glu	Pro	Asp	Ser 155	Ser	Ser	Gly	Ile	Gly 160
Lys	Thr	Gly	Gln	Gln 165	Pro	Ala	Lys	Lys	Arg 170	Leu	Asn	Phe	Gly	Gln 175	Thr
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Ala	Thr	Pro 195	Ala	Ala	Val	Gly	Pro 200	Thr	Thr	Met	Ala	Ser 205	Gly	Gly	Gly
Ala	Pro 210	Met	Ala	Asp	Asn	Asn 215	Glu	Gly	Ala	Asp	G1 y 220	Val	Gly	Asn	Ala
Ser 225	Gly	Asn	Тгр	His	Cys 230	qzA	Ser	Thr	Trp	Leu 235	Gly	Asp	Arg	Val	Ile 240
Thr	Thr	Ser	Thr	Arg	Thr	Trp	Ala	Leu	Pro	Thr	туг	Asn	Asn	His	Leu

				245					250					255	
Tyr	Lys	Gln	11e 260		Ser	Ala	Ser	Thr 265	_	Ala	Ser	Asn	Asp 270		His
Tyr	Phe	Gly 275	_	Ser	Thr	Pro	Trp 280		Туг	Phe	Asp	Phe 285		Arg	Phe
His	Cys 290		Phe	Ser	Pro	Arg 295	-	Trp	Gln	Arg	Leu 300		Asn	Asn	Asn
Trp 305	Gly	Phe	Arg	Pro	Lys 310	-	Leu	Asn	Phe	Lys 315		Phe	Asn	Ile	G1 n 320
Val	Lys	Glu	Val	Thr 325		Asn	Asp	Gly	Val 330		Thr	Ile	Ala	Asn 335	
Leu	Thr	Ser	Thr 340	Val	Gln	Val	Phe	Ser 345		Ser	Glu	Tyr	Gln 350	Leu	Pro
Tyr	Val	Leu 355	Gly	Ser	Ala	His	Gln 360	Gly	Cys	Leu	Pro	Pro 365	Phe	Pro	Ala
Asp	Val 370	Phe	Met	Ile	Pro	Gln 375	Tyr	Gly	Tyr	Leu	Thr 380	Leu	neA	Asn	Gly
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Ser	Gln	Met	Leu	Arg 405	Thr	Gly	Asn	Asn	Phe 410	Thr	Phe	Ser	Tyr	Thr 415	Phe
Glu	Glu	Val	Pro 420	Phe	His	Ser	Ser	Tyr 425	Ala	His	Ser	Gln	Ser 430	Leu	Asp
Arg	Leu	Met 435	Asn	Pro	Leu	Ile	Asp 440	Gln	Tyr	Leu	туг	Tyr 445	Leu	Asn	Arg
	G1n 450				_	455				-	460				
465	Gly				470					475					480
-	Pro			485					490					495	
Asn	Asn	Ser	Asn	Phe	Thr	Trp	Thr	Gly	Ala	Ser	Lys	Tyr	Asn	Leu	Asn

500 505 510

Gly Arg Glu Ser Ile Ile Asn Pro Gly Thr Ala Met Ala Ser His Lys 515 520 525

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Lys Glu Ser Ala Gly Ala Ser Asn Thr Ala Leu Asp Asn Val Met Ile 545 550 555 560

Thr Asp Glu Glu Glu Ile Lys Ala Thr Asn Pro Val Ala Thr Glu Arg 565 570 575

Phe Gly Thr Val Ala Val Asn Phe Gln Ser Ser Ser Thr Asp Pro Ala 580 585 590

Thr Gly Asp Val His Ala Met Gly Ala Leu Pro Gly Met Val Trp Gln 595 600 605

Asp Arg Asp Val Tyr Leu Gln Gly Pro Ile Trp Ala Lys Ile Pro His 610 615 620

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Lys Asn Pro Pro Pro Gln Ile Leu Ile Lys Asn Thr Pro Val Pro Ala 645 650 655

Asn Pro Pro Ala Glu Phe Ser Ala Thr Lys Phe Ala Ser Phe Ile Thr 660 665 670

Gln Tyr Ser Thr Gly Gln Val Ser Val Glu Ile Glu Trp Glu Leu Gln 675 680 685

Lys Glu Asn Ser Lys Arg Trp Asn Pro Glu Val Gln Tyr Thr Ser Asn 690 695 700

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Val Gln Trp Arg Arg Val Ser Lys Ala Pro Glu Ala Leu Phe Phe Val
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acc acg ggg gtc aaa tcc atg gtg ctg ggc cgc ttc ctg agt cag att 336
Thr Thr Gly Val Lys Ser Met Val Leu Gly Arg Phe Leu Ser Gln Ile
100 105 . 110

agg gac aag ctg gtg cag acc atc tac cgc ggg atc gag ccg acc ctg 384 Arg Asp Lys Leu Val Gln Thr Ile Tyr Arg Gly Ile Glu Pro Thr Leu 115 120 125

ccc aac tgg ttc gcg gtg acc aag acg cgt aat ggc gcc gga ggg ggg 433 Pro Asn Trp Phe Ala Val Thr Lys Thr Arg Asn Gly Ala Gly Gly Gly 130 135 140

aac aag gtg gtg gac gag tgc tac atc ccc aac tac ctc ctg ccc aag

Asn Lys Val Val Asp Glu Cys Tyr Ile Pro Asn Tyr Leu Leu Pro Lys

145 150 155 160

act cag ccc gag ctg cag tgg gcg tgg act aac atg gag gag tat ata 528

Thr Gln Pro Glu Leu Gln Trp Ala Trp Thr Asn Met Glu Glu Tyr Ile age gee tgt ttg aac etg gee gag ege aaa egg ete gtg geg eag eac Ser Ala Cys Leu Asn Leu Ala Glu Arg Lys Arg Leu Val Ala Gln His 180 185 ctg acc cac gtc agc cag acc cag gag cag aac aag gag aat ctg aac Leu Thr His Val Ser Gln Thr Gln Glu Gln Asn Lys Glu Asn Leu Asn 195 200 ccc aat tot gac gcg cot gtc atc cgg tea aaa acc tee gcg cgc tac 672 Pro Asn Ser Asp Ala Pro Val Ile Arg Ser Lys Thr Ser Ala Arg Tyr 210 215 220 atg gag ctg gtc ggg tgg ctg gtg gac cgg ggc atc acc tcc gag aag Met Glu Leu Val Gly Trp Leu Val Asp Arg Gly Ile Thr Ser Glu Lys 230 235 cag tgg atc cag gag gac cag gcc tcg tac atc tcc ttc aac gcc gct 768 Gln Trp Ile Gln Glu Asp Gln Ala Ser Tyr Ile Ser Phe Asn Ala Ala 245 250 tee aac teg egg tee eag ate aag gee get etg gae aat gee gge aag 816 Ser Asn Ser Arg Ser Gln Ile Lys Ala Ala Leu Asp Asn Ala Gly Lys 265 atc atg gcg ctg acc aaa tcc gcg ccc gac tac ctg gta ggc ccc gct Ile Met Ala Leu Thr Lys Ser Ala Pro Asp Tyr Leu Val Gly Pro Ala 275 280 285 ccg ccc gcg gac att aaa acc aac cgc atc tac cgc atc ctg gag ctg Pro Pro Ala Asp Ile Lys Thr Asn Arg Ile Tyr Arg Ile Leu Glu Leu 290 295 aac ggc tac gaa cct gcc tac gcc ggc tcc gtc ttt ctc ggc tgg gcc Asn Gly Tyr Glu Pro Ala Tyr Ala Gly Ser Val Phe Leu Gly Trp Ala 310 315 cag aaa agg ttc ggg aag cgc aac acc atc tgg ctg ttt ggg ccg gcc Gln Lys Arg Phe Gly Lys Arg Asn Thr Ile Trp Leu Phe Gly Pro Ala 325 acc acg ggc aag acc aac atc gcg gaa gcc atc gcc cac gcc gtg ccc 1056 Thr Thr Gly Lys Thr Asn Ile Ala Glu Ala Ile Ala His Ala Val Pro 340 345 350 ttc tac ggc tgc gtc aac tgg acc aat gag aac ttt ccc ttc aat gat

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3

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Glu Gln Ala Pro Leu Thr Val Ala Glu Lys Leu Gln Arg Asp Phe Leu 50 55 60

Val Gln Trp Arg Arg Val Ser Lys Ala Pro Glu Ala Leu Phe Phe Val 65 70 75 80

Gln Phe Glu Lys Gly Glu Ser Tyr Phe His Leu His Ile Leu Val Glu 85 90 95

Thr Thr Gly Val Lys Ser Met Val Leu Gly Arg Phe Leu Ser Gln Ile 100 105 110

Arg Asp Lys Leu Val Gln Thr Ile Tyr Arg Gly Ile Glu Pro Thr Leu

120 Pro Asn Trp Phe Ala Val Thr Lys Thr Arg Asn Gly Ala Gly Gly 135 Asn Lys Val Val Asp Glu Cys Tyr Ile Pro Asn Tyr Leu Leu Pro Lys 150 155 Thr Gln Pro Glu Leu Gln Trp Ala Trp Thr Asn Met Glu Glu Tyr Ile 165 170 Ser Ala Cys Leu Asn Leu Ala Glu Arg Lys Arg Leu Val Ala Gln His 185 Leu Thr His Val Ser Gln Thr Gln Glu Gln Asn Lys Glu Asn Leu Asn 200 Pro Asn Ser Asp Ala Pro Val Ile Arg Ser Lys Thr Ser Ala Arg Tyr 215 Met Glu Leu Val Gly Trp Leu Val Asp Arg Gly Ile Thr Ser Glu Lys 235 Gln Trp Ile Gln Glu Asp Gln Ala Ser Tyr Ile Ser Phe Asn Ala Ala 250 Ser Asn Ser Arg Ser Gln Ile Lys Ala Ala Leu Asp Asn Ala Gly Lys Ile Met Ala Leu Thr Lys Ser Ala Pro Asp Tyr Leu Val Gly Pro Ala Pro Pro Ala Asp Ile Lys Thr Asn Arg Ile Tyr Arg Ile Leu Glu Leu Asn Gly Tyr Glu Pro Ala Tyr Ala Gly Ser Val Phe Leu Gly Trp Ala Gln Lys Arg Phe Gly Lys Arg Asn Thr Ile Trp Leu Phe Gly Pro Ala 330 Thr Thr Gly Lys Thr Asn Ile Ala Glu Ala Ile Ala His Ala Val Pro 340 345 Phe Tyr Gly Cys Val Asn Trp Thr Asn Glu Asn Phe Pro Phe Asn Asp 360

Cys Val Asp Lys Met Val Ile Trp Trp Glu Glu Gly Lys Met Thr Ala

•	70 00	12000	•												• •
	370					375					380				
Lys 385	Val	Val	G1u	Ser	Ala 390	Lys	Ala	Ile	Leu	Gly 395	Gly	Ser	Lys	Val	Arç
Val	Asp	Gln	Lys	Cys 405	Lys	Ser	Ser	Ala	Gln 410	Ile	Asp	Pro	Thr	Pro 415	
Ile	Val	Thr	Ser 420	Asn	Thr	Asn	Met	Cys 425		Val	Ile	Asp	Gly 430	Asn	Ser
Thr	Thr	Phe 435	Glu	His	Gln	Gln	Pro 440	Leu	Gln	Asp	Arg	Met 445	Phe	Lys	Phe
Glu	Leu 450	Thr	_	Arg	Leu	Glu 455	His	Asp	Phe	Gly	Lys 460	Val	Thr	Lys	Glr
Glu 465	Val	Lys	Glu	Phe	Phe 470	Arg	Trp	Ala	Gln	Asp 475	His	Val	Thr	G1u	Val 480
Ala	His	Glu	Phe	Tyr 485	Val	Arg	Lys	Gly	Gly 490	Ala	Asn	Lys	Arg	Pro 495	Ala
Pro	Asp	Asp	Ala 500	Asp	Lys	Ser	Glu	Pro 505	Lys	Arg	Ala	Cys	Pro 510	Ser	Val
Ala	Asp	Pro 515	Ser	Thr	Ser	Asp	Ala 520	Glu	Gly	Ala	Pro	Val 525	Asp	Phe	Ala
Asp	Arg 530	Туг	Gln	Asn	Lys	Cys 535	Ser	Arg	His	Ala	Gly 540	Met	Leu	Gln	Met
Leu 545	Phe	Pro	Cys	Lys	Thr 550	Cys	Glu	Arg	Met	Asn 555	Gln	Asn	Phe	Asn	Ile 560
Cys	Phe	Thr	His	Gly 565	Thr	Arg	Asp	Cys	Ser 570	Glu	Cys	Phe	Pro	Gly 575	Val
Ser	Glu	Ser	Gln 580	Pro	Val	Val	Arg	Lys 585	Arg	Thr	Tyr	Arg	Lys 590	Leu	Cys
Ala	Ile	His 595	His	Leu	Leu	Gly	Arg 600	Ala	Pro	Glu	Ile	Ala 605	Cys	Ser	Ala

Cys Asp Leu Val Asn Val Asp Leu Asp Asp Cys Val Ser Glu Gln 610 620

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,	wo 0	0/280	61												PC	T/US99/25694	
145	•				150)				155	5				160		
	-		-		Glr					Ası				•	t ata Tle	528	
				ı Asr					Lys					Glr	g cac n His	576	
			Val					Glu					Asn		aac Asn	624	
		Ser	-				Ile					Ser		-	tac Tyr	672	
	Glu				Trp 230	Leu					Ile				aag Lys 240	720	
					gac Asp									-	Ala	768	
				Ser	cag Gln					-	-		-		-	816	
					aaa Lys						-	-			-	864	
					aaa Lys								-		_	912	
					gcc Ala 310					_					•	960	
					aag Lys						_			-	_	1008	
					aac Asn											1056	

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ttc	tac	ggc	tgc	gtc	aac	tgg	acc	aat	gag	aac	ttt	ccc	ttc	aat	gat	1104
					Asn											
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_	-	-	_	_	gtg											1152
Cys		Asp	Lys	Met	Val		Trp	Trp	Glu	Glu		Lys	Met	Thr	Ala	
	370					375					380					
																1000
•	-				gcc											1200
-	Val	Val	GIU	ser	Ala 390	Lys	Ala	ite	Leu	395	GIY	261	гуя	val	400	
385					390					333					400	
ata	cac	caa	aan	tac	aag	tca	tcc	acc	cad	atc	gac	ccc	acc	ccc	ata	1248
					Lys											
VGI	م س	01.	2,0	405	2,2	-			410					415		
atc	qtc	acc	tcc	aac	acc	aac	atg	tgc	gcc	gtg	att	gac	ggg	aac	agc	1296
	-				Thr											
			420					425					430			
acc	acc	ttc	gag	cac	cag	cag	ccg	ttg	cag	gac	cgg	atg	ttc	aaa	ttt	1344
Thr	Thr	Phe	Glu	His	Gln	Gln	Pro	Leu	Gln	Asp	Arg		Phe	Lys	Phe	
		435					440					445				
-					ctg											1392
Glu		Thr	Arg	Arg	Leu		His	Asp	Phe	GIÀ		Val	Thr	Lys	Gin	
	450					455					460					
~~~	a+ a		a 2 a	++6	ttc		tric	aca	can	gat	cac	ata	acc	nan	ata	1440
-	-				Phe	-			_	-						1
465	741	Lyo	014		470	,,r. g		,,,,,,		475					480	
qcq	cat	qaq	ttc	tac	gtc	aga	aag	ggt	gga	gcc	aac	aaa	aga	ccc	gcc	1488
					Val											
				485					490					495		
ccc	gat	gac	gcg	-	aaa	•			-		-	-				1536
n		T		T		C	C1	D	* ***	T	81.	C	Dra	C ~ ~	17 - 1	

Pro Asp Asp Ala Asp Lys Ser Glu Pro Lys Arg Ala Cys Pro Ser Val

geg gat cca teg acg tea gae geg gaa gga get eeg gtg gae ttt gee Ala Asp Pro Ser Thr Ser Asp Ala Glu Gly Ala Pro Val Asp Phe Ala 520

gac agg tat ggc tgc cga tgg tta tct tcc aga ttg gct cga gga caa

Asp Arg Tyr Gly Cys Arg Trp Leu Ser Ser Arg Leu Ala Arg Gly Gln

1632

505

500

WO 00/28061	PCT/US99/25694
WO 00/28061	PC1/US99/25094

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cct ctc tga 1641 Pro Leu

Pro Leu 545

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<213> AAV-1

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Glu His Leu Pro Gly Ile Ser Asp Ser Phe Val Ser Trp Val Ala Glu 20 25 30

Lys Glu Trp Glu Leu Pro Pro Asp Ser Asp Met Asp Leu Asn Leu Ile  $35 \hspace{1cm} 40 \hspace{1cm} 45 \hspace{1cm}$ 

Glu Gln Ala Pro Leu Thr Val Ala Glu Lys Leu Gln Arg Asp Phe Leu 50 55 60

Val Gln Trp Arg Arg Val Ser Lys Ala Pro Glu Ala Leu Phe Phe Val 65 70 75 80

Gln Phe Glu Lys Gly Glu Ser Tyr Phe His Leu His Ile Leu Val Glu 85 90 95

Thr Thr Gly Val Lys Ser Met Val Leu Gly Arg Phe Leu Ser Gln Ile 100 105 110

Arg Asp Lys Leu Val Gln Thr Ile Tyr Arg Gly Ile Glu Pro Thr Leu 115 120 125

Pro Asn Trp Phe Ala Val Thr Lys Thr Arg Asn Gly Ala Gly Gly Gly 130 135 140

Asn Lys Val Val Asp Glu Cys Tyr Ile Pro Asn Tyr Leu Leu Pro Lys 145 150 155 160

Thr Gln Pro Glu Leu Gln Trp Ala Trp Thr Asn Met Glu Glu Tyr Ile 165 170 170 175

Ser Ala Cys Leu Asn Leu Ala Glu Arg Lys Arg Leu Val Ala Gln His 180 185 190

Leu Thr His Val Ser Gln Thr Gln Glu Gln Asn Lys Glu Asn Leu Asn Pro Asn Ser Asp Ala Pro Val Ile Arg Ser Lys Thr Ser Ala Arg Tyr 215 Met Glu Leu Val Gly Trp Leu Val Asp Arg Gly Ile Thr Ser Glu Lys 230 Gln Trp Ile Gln Glu Asp Gln Ala Ser Tyr Ile Ser Phe Asn Ala Ala 245 250 Ser Asn Ser Arg Ser Gln Ile Lys Ala Ala Leu Asp Asn Ala Gly Lys 265 Ile Met Ala Leu Thr Lys Ser Ala Pro Asp Tyr Leu Val Gly Pro Ala 280 Pro Pro Ala Asp lle Lys Thr Asn Arg Ile Tyr Arg Ile Leu Glu Leu Asn Gly Tyr Glu Pro Ala Tyr Ala Gly Ser Val Phe Leu Gly Trp Ala 315 Gln Lys Arg Phe Gly Lys Arg Asn Thr Ile Trp Leu Phe Gly Pro Ala 330 Thr Thr Gly Lys Thr Asn Ile Ala Glu Ala Ile Ala His Ala Val Pro 345 Phe Tyr Gly Cys Val Asn Trp Thr Asn Glu Asn Phe Pro Phe Asn Asp 360 Cys Val Asp Lys Met Val Ile Trp Trp Glu Glu Gly Lys Met Thr Ala 375 Lys Val Val Glu Ser Ala Lys Ala Ile Leu Gly Gly Ser Lys Val Arg 390 395 Val Asp Gln Lys Cys Lys Ser Ser Ala Gln Ile Asp Pro Thr Pro Val 410

Ile Val Thr Ser Asn Thr Asn Met Cys Ala Val Ile Asp Gly Asn Ser

Thr Thr Phe Glu His Gln Gln Pro Leu Gln Asp Arg Met Phe Lys Phe 435 440 445

Glu Leu Thr Arg Arg Leu Glu His Asp Phe Gly Lys Val Thr Lys Gln

Glu Val Lys Glu Phe Phe Arg Trp Ala Gln Asp His Val Thr Glu Val 470 475 Ala His Glu Phe Tyr Val Arg Lys Gly Gly Ala Asn Lys Arg Pro Ala 490 Pro Asp Asp Ala Asp Lys Ser Glu Pro Lys Arg Ala Cys Pro Ser Val 505 Ala Asp Pro Ser Thr Ser Asp Ala Glu Gly Ala Pro Val Asp Phe Ala 520 Asp Arg Tyr Gly Cys Arg Trp Leu Ser Ser Arg Leu Ala Arg Gly Gln 535 540 Pro Leu 545 <210> 8 <211> 1200 <212> DNA <213> AAV-1 <220> <221> CDS <222> (1)..(1197) atg gag ctg gtc ggg tgg ctg gtg gac cgg ggc atc acc tcc gag aag Met Glu Leu Val Gly Trp Leu Val Asp Arg Gly Ile Thr Ser Glu Lys cag tgg atc cag gag gac cag gcc tcg tac atc tcc ttc aac gcc gct Gln Trp Ile Gln Glu Asp Gln Ala Ser Tyr Ile Ser Phe Asn Ala Ala 20 25 tee aac teg egg tee eag ate aag gee get etg gae aat gee gge aag Ser Asn Ser Arg Ser Gln Ile Lys Ala Ala Leu Asp Asn Ala Gly Lys 40 atc atg gcg ctg acc aaa tcc gcg ccc gac tac ctg gta ggc ccc gct Ile Met Ala Leu Thr Lys Ser Ala Pro Asp Tyr Leu Val Gly Pro Ala 60 · 55

PCT/US99/25694 WO 00/28061

		gcg														240
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65					70					75					80	
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Asn	G1 v	Tyr	Glu	Pro	Ala	Tyr	Ala	Gly	Ser	Val	Phe	Leu	Gly	Trp	Ala	
	2			85		-		•	90				_	95		
cag	aaa	agg	ttc	ggg	aag	cgc	aac	acc	atc	tgg	ctg	ttt	ggg	ccg	gcc	336
Gln	Lys	Arg		Gly	Lys	Arg	Asn		Ile	тгр	Leu	Phe		Pro	Ala	
			100					105					110			
		ggc		200	226	250	aca	~~~	acc	atc	acc	cac	acc	ata	ccc	384
acc	acg Thr	Gly	Lve	Thr	Asn	Tie	Ala	Glu	Ala	Ile	Ala	His	Ala	Val	Pro	
III	1111	115	Dys	****	7131.		120					125				
ttc	tac	ggc	tgc	gtc	aac	tgg	acc	aat	gag	aac	ttt	ccc	ttc	aat	gat	432
Phe	Tyr	Gly	Cys	Val	Asn		Thr	Asn	Glu	Asn		Pro	Phe	Asn	Asp	
	130					135					140					
		gac	224	250	ata	atc	Faa	rog	aaa	gag	aac	aag	ato	aco	acc	480
		Asp														
145	•	пор	-,-		150		•	•		155	•	•			160	
aag	gtc	gtg	gag	tcc	gcc	aag	gcc	att	ctc	ggc	ggc	agc	aag	gtg	cgc	528
Lys	Val	Val	Glu		Ala	Lys	Ala	Ile		Gly	Gly	Ser	Lys		Arg	
				165					170					175		
~+~	~~~	caa	220	t a c	220	tca	tcc	acc	can	atc	gac	ccc	acc	ccc	ata	576
		Gln														
			180	-,-	-,-			185			•		190			
		acc														624
Ile	Val	Thr	Ser	Asn	Thr	Asn		Cys	Ala	Val	Ile		Gly	Asn	Ser	
		195					200					205				
200	200	ttc	<b>~~~</b>	C 3 C	can	can	cca	tta	cag	gac	caa	atσ	ttc	aaa	ttt	672
		Phe														
	210					215				-	220			•		
gaa	ctc	acc	cgc	cgt	ctg	gag	cat	gac	ttt	ggc	aag	gtg	aca	aag	cag	720
	Leu	Thr	Arg	Arg		Glu	His	Asp	Phe		Lys	Val	Thr	Lys		
225					230					235					240	
(12.2)	atc	aaa	aaa	ttc	tte	cac	Laa	aca	caa	gat	cac	ata	acc	gag	ata	768
		Lys														
		-,-		245		,	•		250	•				255		
							27									

gcg	cat	ga	g tt	c ta	c gto	aga	a aag	ggt	t gga	gco	aa	сааа	aga	c c	c gcc	816
Ala	1 H18	Gl	u Ph: 26	е Ту. О	r Val	l Arç	Lys	Gly 265		Ala	Ası	n Lys	270		o Ala	
CCC	gat	gad	ge	g gai	t aaa	ago	gag	ccc	aag	cgg	gco	: tgc	ccc	: tc	a gtc	864
Pro	ASP	275	P ATS	a As	Lys	Ser	Glu 260		Lys	Arg	Ala	285 Cys		Se	r Val	
gcg	gat	CC	tcc	aco	tca	gac	gcg	gaa	gga	gct	ccg	gtg	gac	tt	t gcc	912
AId	290	PIC	) Sei	Thi	Ser	295	Ala	Glu	Gly	Ala	9ro 300		Asp	Phe	≥ Ala	
gac	agg	tac	caa	aac	aaa	tgt	tct	cgt	cac	gcg	ggc	atg	ctt	caç	gatg	960
305	ALY	ıyı	GII	ASI	310	Cys	ser	Arg	His	A1a 315	Gly	Met	Leu	Glr	Met 320	
ctg	ttt	CCC	tgc	aag	aca	tgc	gag	aga	atg	aat	cag	aat	ttc	aac	att	1008
			Cys	325		Cys	GIU	Arg	330	ASN	Gin	Asn	Phe	335		
tgc Cvs	ttc Phe	acg Thr	Cac	999 G1 v	acg	aga	gac	tgt	tca	gag	tgc	ttc Phe	ccc	ggc	gtg	1056
-,-		••••	340	Ory	****	ALG	nsp	345	ser	GIU	cys	Pne	350	Gly	Val	
Ser	gaa Glu	Ser	Caa Gln	CCG Pro	gtc Val	gtc Val	aga	aag	agg	acg	tat	cgg Arg	aaa	ctc	tgt	1104
		355					360	<b>_</b> y	ALG	1111	1 y L	365	rys	Leu	Cys	
~~~																
Ala	Ile	His	His	Leu	Leu	ggg Glv	cgg Ara .	gct Ala	Pro	gag Glu	att	gct Ala	tgc	tcg	gcc	1152
	370					375			•••		380	<i>_</i>	cys	Set	ALA	
tac	a = t	a.	~+~													
Cys .	Asp	Leu	Val	Asn	Val.	gac Asp	Leu 1	gat Asp i	gac (Asp (tgt : Cvs :	gtt Val	tct o	gag	caa Gln	taa	1200
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														15		
oln 1	rp 1	le (3ln (3lu A	Asp G	ln A	la s	er T	'yr I	le S	er E	he A	sn A	la i	Ala	

		, w 200	•••												•
			20)				25	•				30)	
Ser	Asn	Ser 35	-	Ser	Gln	Ile	Lys 40		Ala	Leu	Asp	Asn 45		Gl y	Lys
Ile	Met 50		Leu	Thr	Lys	Ser 55		Pro	Asp	Туг	Leu 60		Gly	Pro	Ala
Pro 65	Pro	Ala	Asp	Ile	Lys 70		Asn	Arg	Ile	Tyr 75	-	Ile	Leu	Glu	Leu 80
Asn	Gly	Tyr	Glu	Pro 85		Tyr	Ala	Gly	Ser 90		Phe	Leu	Gly	Trp 95	
Gln	Lys	Arg	Phe 100	Gly	Lys	Arg	Asn	Thr 105		Trp	Leu	Phe	Gly 110		Ala
Thr	Thr	Gly 115	Lys	Thr	Asn	Ile	Ala 120	Glu	Ala	Ile	Ala	His 125	Ala	Val	Pro
Phe	Tyr 130	Gly	Суз	Val	Asn	Trp 135	Thr	Asn	Glu	Asn	Phe 140	Pro	Phe	Asn	Asp
Cys 145	Val	Asp	Lys	Met	Val 150	Ile	Тгр	Trp	Glu	Glu 155	Gly	Lys	Met	Thr	Ala 160
Lys	Val	Val	Glu	Ser 165	Ala	Lys	Ala	Ile	Leu 170	Gly	Gly	ser	Lys	Val 175	Arg
Val	Asp	Gln	Lys 180	Cys	Lys	Ser	Ser	Ala 185	Gln	Ile	Asp	Pro	Thr 190	Pro	Val
Ile	Val	Thr 195	Ser	Asr	Thr	Asn	Met 200	Cys	Ala	Val	Ile	Asp 205	Gly	Asn	Ser
Thr	Thr 210	Phe	Glu	His	Gln	Gln 215	Pro	Leu	Gln	Asp	Arg 220	Met	Phe	Lys	Phe
G1u 225	Leu	Thr	Arg	Arg	Leu 230	Glu	His	Asp	Phe	Gly 235	Lys	Val	Thr	Lys	Gln 240
Glu	Val	Lys	Glu	Phe 245	Phe	Arg	Trp	Ala	G1n 250	Asp	His	Val	Thr	Glu 255	Val
Ala	His	Glu	Phe	Туr	Val	Arg	Lys	Gly	Gly	Ala	Asn	Lys	Arg	Pro	Ala

265 Pro Asp Asp Ala Asp Lys Ser Glu Pro Lys Arg Ala Cys Pro Ser Val

285

275 280

Ala Asp Pro Ser Thr Ser Asp Ala Glu Gly Ala Pro Val Asp Phe Ala 290 295 300

Asp Arg Tyr Gln Asn Lys Cys Ser Arg His Ala Gly Met Leu Gln Met 305 310 315 320

Leu Phe Pro Cys Lys Thr Cys Glu Arg Met Asn Gln Asn Phe Asn Ile 325 330 335

Cys Phe Thr His Gly Thr Arg Asp Cys Ser Glu Cys Phe Pro Gly Val 340 345 350

Ser Glu Ser Gln Pro Val Val Arg Lys Arg Thr Tyr Arg Lys Leu Cys 355 360 365

Ala Ile His His Leu Leu Gly Arg Ala Pro Glu Ile Ala Cys Ser Ala 370 375 380

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<221> misc feature

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1 5 10 15

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tcc aac tcg cgg tcc cag atc aag gcc gct ctg gac aat gcc ggc aag 144

	WUU	W280	01												r	. 170377/4
Ser	Asn	Ser 35	-	Ser	Gln	Ile	Lys 40		Ala	Leu	Asp	Asn 45	Ala	Gly	Lys	
	-		-						_		_	_			gct Ala	192
-	Pro		-			Thr		-			Arg		_		ctg Leu 80	240
	ggc Gly		-		-		-			-					-	288
_	aaa Lys															336
	acg Thr															384
	tac Tyr 130		_	_											_	432
-	gtc Val	-	-									-	-	-	-	480
_	gtc Val				-							-	_		-	528
	gac Asp							_	_		-				-	576
	gtc Val					Asn									-	624
Thr	acc Thr 210		-		-	-	-	-	-	-		_				672
gaa	ctc	acc	cgc	cgt	ctg	gag	cat	gac	ttt	ggc	aag	gtg	aca	aag	cag	720

,	VO 0	0/2800	51												РСТ	/US99/25694
Glu 225	Leu	Thr	Arg	Arg	Leu 230		His	Asp	Phe	Gly 235	Lys	Val	Thr	Lys	Gln 240	
•	-	aaa Lys			Phe	_										768
		gag Glu														816
	-	gac Asp 275		-		-			-		-	-			-	864
	-	cca Pro	-	-		-	-	-		-			-		-	912
-		tat Tyr		-	-					-	-	-	-			960
cct Pro	ctc Leu	tga														969
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Ser	Asn	ser 35	Arg	Ser	Gln	Ile	Lys 40	Ala	Ala	Leu	Asp	Asn 45	Ala	Gly	Lys	
Ile	Met 50	Ala	Leu	Thr	Lys	Ser 55	Ala	Pro	Asp	Tyr	Leu 60	Val	Gly	Pro	Ala	
Pro 65	Pro	Ala	Asp	Ile	Lys 70	Thr	Asn	Arg	lle	Туг 75	Arg	Ile	Leu	Glu	Leu 80	

Asn	Gly	Tyr	Glu	Pro 85		Tyr	: Ala	Gly	Ser 90		Phe	. Lev	ı Gly	Trp 95	
Gln	Lys	Arg	Phe 100	-	Lys	Arg) Asn	Thr 105		Trp	Leu	Phe	Gly		Ala
Thr	Thr	Gly 115	Lys	Thr	Asn	Ile	120		Ala	Ile	Ala	His 125		Val	Pro
Phe	Туг 130	-	Cys	Val	Asn	Trp		Asn	Glu	Asn	Phe 140		Phe	Asn	Asp
Cys 145		Asp	Lys	Met	Val 150		Тгр	Trp	Glu	Glu 155	_	Lys	Met	Thr	Ala 160
Lys	Val	Val	Glu	Ser 165		Lys	Ala	Ile	Leu 170	_	G1 y	Ser	Lys	Val 175	-
Val	Asp	Gln	Lys 180	Cys	Lys	Ser	Ser	Ala 185	Gln	Ile	Asp	Pro	Thr 190	Pro	Val
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Ala	Asp 290	Pro	Ser	Thr	Ser	Asp 295	Ala	Glu	Gly	Ala	Pro 300	Val	Asp	Phe	Ala
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Pro Leu

305

320

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Glu	GLY	11e	Arg 20		Trp	Trp	Asp	Leu 25		PFO	GIA	ALA	30		Pro	
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Gln	Gln	Leu	Lys		Gly	Asp	Asn	Pro		Leu	Arg	Tyr	Asn		Ala	
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Leu	-	Leu	Val	Glu	Glu	-	Ala	Lys	Thr	Ala		Gly	Lys	Lys	Arg	
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					acg Thr			Gly								1008
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gaq Gli	g g	aa lu '	gtg Val	ect Pro 420	Pho	с са е Н:	ac a is S	gc :	ser	tac Tyr 42 5	gcg Ala	cac His	agc Ser	Gln :	agc (Ser 1	etg Leu	gac Asp	1296
cgg Arg	C1	su r	atg Met 135	aat Asn	Pro	ct Le	c a	le F	Jac Asp	caa Gln	tac Tyr	ctg Leu	tat i	tac c Fyr I 145	tg a Jeu A	ac a	aga Arg	1344
act Thr	G1 45	.11 М	at Isn	cag Gln	tcc Ser	gg G1	a aq y Se 45	er A	cc d	aa . Sln /	aac Asn :	Lys .	gac t Asp L 460	tg c	tg t eu P	tt a	igc ier	1392
cgt Arg 465	gg G1	g t y S	ct o	cca Pro	gct Ala	gg(G1) 47(y Me	g to	ct g er V	tt d	31n Z	ecc a Pro I	aaa a Lys A	ac to	gg c	eu P	ct ro 80	1440
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ggg Gly	egt Arg	ga G1 51	u 50	cc a	itc le	atc Ile	aac Asn	52	c GI	y Th	et go	ct as	tg gc et Al 52	a Se	a ca r Hi:	c aa s Ly	a 's	1584
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t t Ph	t g ie G	gg ly	acc Thr	gtg Val 580	gca	gta Val	aat Asn	ttc Phe	cag Gln 585	agc Ser	agc Ser	agc Ser	Thr .	gac Asp 590	cct Pro	gcg Ala	1776
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Thi	r As	p G	ga (cac	ttt Phe	cac His 630	ccg Pro :	tct (cct Pro	Leu	atg (Met (635	ggc g Gly G	ggc t	tt o	31 y	ctc Leu 640	1920
aaç Lys	aa As	c c n P	cg o	ro	cct Pro 645	cag Gln	atc o	etc a	le.	aaa Lys i 650	aac a Asn T	ncg c	ct g	al P	ct ro	gcg Ala	1968
aat Asn	Pro	t co	ro A	cg (la (gag Glu	ttt Phe	tca g Ser A	la T	ca a hr 1 65	aag t Lys I	tt g	ct to	ca ti er Pl 67	ne I	tc a	ecc Thr	2016
caa Gln	tac Tyr	5 to 5 Se 67	rT	ca ç hr G	ga (caa (Gln '	gtg a Val S 6	gt g er V 80	ty g	aa a lu I	tt g	aa to lu Ti 68	rp G1	ng ci	tg c	ag 11n	2064
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tat Tyr 705	gca Ala	aa Ly	a to s Se	et g	la A	ac g sn V	tt ga al As	nt to Sp Ph	t a	ct gr hr Va 7:	al As	ic aa ip As	c aa n As:	t gg n Gl	y L	tt eu 20	2160
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Gly Tyr Lys Tyr Leu Gly Pro Phe Asn Gly Leu Asp Lys Gly Glu Pro 50 55 60

Val Asn Ala Ala Asp Ala Ala Ala Leu Glu His Asp Lys Ala Tyr Asp 65 70 75 80

Gln Gln Leu Lys Ala Gly Asp Asn Pro Tyr Leu Arg Tyr Asn His Ala 85 90 95

Asp Ala Glu Phe Gln Glu Arg Leu Gln Glu Asp Thr Ser Phe Gly Gly 100 105 110

As Leu Gly Arg Ala Val Phe Gln Ala Lys Lys Arg Val Leu Glu Pro 115 120 125

Leu Gly Leu Val Glu Glu Gly Ala Lys Thr Ala Pro Gly Lys Lys Arg 130 135 140

Pro Val Glu Gln Ser Pro Gln Glu Pro Asp Ser Ser Ser Gly Ile Gly 145 150 155 160

Lys Thr Gly Gln Gln Pro Ala Lys Lys Arg Leu Asn Phe Gly Gln Thr 165 170 175

Gly Asp Ser Glu Ser Val Pro Asp Pro Gln Pro Leu Gly Glu Pro Pro 180 185 190

Ala Thr Pro Ala Ala Val Gly Pro Thr Thr Met Ala Ser Gly Gly Gly 195 200 205

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- Val Lys Glu Val Thr Thr Asn Asp Gly Val Thr Thr Ile Ala Asn Asn 325 330 335
- Leu Thr Ser Thr Val Gln Val Phe Ser Asp Ser Glu Tyr Gln Leu Pro 340 345
- Tyr Val Leu Gly Ser Ala His Gln Gly Cys Leu Pro Pro Phe Pro Ala 355 360 365
- Asp Val Phe Met Ile Pro Gln Tyr Gly Tyr Leu Thr Leu Asn Asn Gly
- Ser Gln Ala Val Gly Arg Ser Ser Phe Tyr Cys Leu Glu Tyr Phe Pro 385 390 395 400
- Ser Gln Met Leu Arg Thr Gly Asn Asn Phe Thr Phe Ser Tyr Thr Phe
 405 410 415
- Glu Glu Val Pro Phe His Ser Ser Tyr Ala His Ser Gln Ser Leu Asp 420 425 430
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Thr Gln Asn Gln Ser Gly Ser Ala Gln Asn Lys Asp Leu Leu Phe Ser 450 460

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- Gly Pro Cys Tyr Arg Gln Gln Arg Val Ser Lys Thr Lys Thr Asp Asn 485 490 495
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- Gly Arg Glu Ser Ile Ile Asn Pro Gly Thr Ala Met Ala Ser His Lys $515 \hspace{1.5cm} 520 \hspace{1.5cm} 525 \hspace{1.5cm}$
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- Lys Glu Ser Ala Gly Ala Ser Asn Thr Ala Leu Asp Asn Val Met Ile 545 550 555 560
- Thr Asp Glu Glu Glu Ile Lys Ala Thr Asn Pro Val Ala Thr Glu Arg 565 570 575
- Phe Gly Thr Val Ala Val Asn Phe Gln Ser Ser Ser Thr Asp Pro Ala 580 585 550
- Thr Gly Asp Val His Ala Met Gly Ala Leu Pro Gly Met Val Trp Gln 595 600 605
- Asp Arg Asp Val Tyr Leu Gln Gly Pro Ile Trp Ala Lys Ile Pro His 610 615 620
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- Asn Pro Pro Ala Glu Phe Ser Ala Thr Lys Phe Ala Ser Phe Ile Thr 660 665 670
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Pro Thr Tyr Asn Asn His Leu Tyr Lys Gln Ile Ser Ser Ala Ser Thr
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Tyr	Phe	Asp	Phe	Asn	Arg	Phe	His	Cys	His	Phe	Ser	Pro	Arg	Asp	Trp	
145					150					155					160	
		-t-			aac		+ = =	~~~	++-		ccc	220	272	ctc	220	528
-	_				Asn											320
GIII	ALG	Deu	116	165	A311	۸۵.	110	01,	170	7129	•••	2,0	,,,,	175		
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Phe	Lys	Leu		Asn	Ile	Gln	Val	_	Glu	Val	Thr	Thr		Asp	Gly	
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Val	Thr	Thr	Ile	Ala	Asn	Asn	Leu	Thr	Ser	Thr	Val	Gln	Val	Phe	Ser	
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-	-			_	Leu	-										
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Суз	Leu	Pro	Pro	Phe	Pro	Ala	Asp	Val	Phe	Met	Ile	Pro	Gln	Tyr	Gly	
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Phe	Thr	Phe	Ser	Tyr	Thr	Phe	Glu	Glu	Val	Pro	Phe	His	Ser	Ser	Tyr	
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Ası	ı Lys	s As	p Le	1 Let 32:		e Ser	Arq	g G1			o Al	a Gl	у Ме		r Val	
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**						- 220									t ggt	1104
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Thr 385		Met	Ala	Ser			Asp	Asp	Glu			Phe	Phe	Pro	Met	
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			gcc Ala													1344
		435		••••	oru	9	440	OI y		Val	A1 a	445	ASII	Pile	GIII	
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ser	450	ser	Thr	Asp	Pro	A1A 455	Thr	GIy	Asp	Val	His 460	Ala	Met	Gly	Ala	
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			Lys													
				485					490					495		
-++	ato	aac	aac		aa 3	cto									- •	
Leu	Met	Gly	ggc Gly	Phe	gya Glv	Leu I	aag Lys .	aac Asn	Pro	Pro	Pro	Gln	atc Ile	Lev	atc	1536
		•	500		- •			505					510	J-04	-10	

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85	90	95

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- Gln Arg Leu Ile Asn Asn Trp Gly Phe Arg Pro Lys Arg Leu Asn 165 170 175
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- Val Thr Thr Ile Ala Asn Asn Leu Thr Ser Thr Val Gln Val Phe Ser 195 200 205
- Asp Ser Glu Tyr Gln Leu Pro Tyr Val Leu Gly Ser Ala His Gln Gly 210 215 220
- Cys Leu Pro Pro Phe Pro Ala Asp Val Phe Met Ile Pro Gln Tyr Gly 225 230 235 240
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- Ala His Ser Gln Ser Leu Asp Arg Leu Met Asn Pro Leu Ile Asp Gln 290 295 300
- Tyr Leu Tyr Tyr Leu Asn Arg Thr Gln Asn Gln Ser Gly Ser Ala Gln 305 310 315 320
- Asn Lys Asp Leu Leu Phe Ser Arg Gly Ser Pro Ala Gly Met Ser Val 325 330 335
- Gln Pro Lys Asn Trp Leu Pro Gly Pro Cys Tyr Arg Gln Gln Arg Val

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Ala Ser Lys Tyr Asn Leu Asn Gly Arg Glu Ser Ile Ile Asn Pro Gly 370 375 380

Thr Ala Met Ala Ser His Lys Asp Asp Glu Asp Lys Phe Phe Pro Met 385 390 395 400

Ser Gly Val Met Ile Phe Gly Lys Glu Ser Ala Gly Ala Ser Asn Thr 405 410 415

Ala Leu Asp Asn Val Met Ile Thr Asp Glu Glu Glu Ile Lys Ala Thr 420 425 430

Asn Pro Val Ala Thr Glu Arg Phe Gly Thr Val Ala Val Asn Phe Gln 435 440 445

Ser Ser Ser Thr Asp Pro Ala Thr Gly Asp Val His Ala Met Gly Ala 450 455 460

Leu Pro Gly Met Val Trp Gln Asp Arg Asp Val Tyr Leu Gln Gly Pro 465 470 475 480

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Leu Met Gly Gly Phe Gly Leu Lys Asn Pro Pro Pro Gln Ile Leu Ile 500 505 510

Lys Asn Thr Pro Val Pro Ala Asn Pro Pro Ala Glu Phe Ser Ala Thr 515 520 525

Lys Phe Ala Ser Phe Ile Thr Gln Tyr Ser Thr Gly Gln Val Ser Val 530 535 540

Glu Ile Glu Trp Glu Leu Gln Lys Glu Asn Ser Lys Arg Trp Asn Pro 545 550 . 555 560

Glu Val Gln Tyr Thr Ser Asn Tyr Ala Lys Ser Ala Asn Val Asp Phe 565 570 575

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Ser 145	G11	j ta i Ty	c ca	g ct n Le	t cc u Pro 15	о Туі	gto Val	c ct	c ggo u Gly	Sei 15:	r Ala	g cada	c ca s Gl	g gg n Gl	c tgc y Cys 160	480
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				n Ası					val					r Ph	t tac e Tyr	576
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Ala Ser Asn Asp Asn His Tyr Phe Gly Tyr Ser Thr Pro Trp Gly Tyr 65 70 75 80

Phe Asp Phe Asn Arg Phe His Cys His Phe Ser Pro Arg Asp Trp Gln 85 90 95

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Lys Leu Phe Asn Ile Gln Val Lys Glu Val Thr Thr Asn Asp Gly Val 115 120 125

Thr Thr Ile Ala Asn Asn Leu Thr Ser Thr Val Gln Val Phe Ser Asp 130 135 140

Ser Glu Tyr Gln Leu Pro Tyr Val Leu Gly Ser Ala His Gln Gly Cys 145 150 155 160

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- Leu Tyr Tyr Leu Asn Arg Thr Gln Asn Gln Ser Gly Ser Ala Gln Asn 245 250 255
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- Pro Gly Met Val Trp Gln Asp Arg Asp Val Tyr Leu Gln Gly Pro Ile 405 410 415
- Trp Ala Lys Ile Pro His Thr Asp Gly His Phe His Pro Ser Pro Leu 420 425 430
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Phe Ala Ser Phe Ile Thr Gln Tyr Ser Thr Gly Gln Val Ser Val Glu 465 470 475 480

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INTERNATIONAL SEARCH REPORT

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C. DOCUM	ENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the re	levant passages	Relevant to claim No.
Y	RUTLEDGE E. A. ET AL.: "Infecti and vectors derived from adeno-a virus (AAV) serotypes other than	ssociated	1-23
	2." JOURNAL OF VIROLOGY, vol. 72, no. 1, January 1998 (19) pages 309-319, XP002137089 ISSN: 0022-538X cited in the application the whole document	98-01),	
Y	WO 98 11244 A (SAFER BRIAN ;US HI (US); CHIORINI JOHN A (US); KOTII M) 19 March 1998 (1998–03–19) the whole document	EALTH N ROBERT -/	1-23
X Furth	er documents are listed in the construction of box C.	X Patent family members are Sated	in erner
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"A" docume consider the earlier of filing de "L" documes which is cluston "O" documes other m"?" documes "P" docum	nt which may throw doubts on priority claim(s) or a cited to establish the publication date of arrother or other speciel reason (se specified) nt referring to an oral disclosure, use, achibition or	T' later document published after the inter- or priority date and not in conflict with- cited to understand the principle or the invention "X" document of particular relevance; the o- cannot be considered novel or cesnot involve an inventive step when the do- "Y" document of particular relevance; the of- cannot be considered to involve an in- document is combined with one or mo- ments, such combination being obviou- in the ext. "A" document member of the seme patent if	idinal invention be considered to be considered to cument is taken alone simed invention entities exp when the re other such docu- is to a person addied
Date of the e	ctual completion of the international search	Outs of mailing of the international sea	rch report
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Name and m	alling address of the ISA Europeen Pettent Office, P.B. 6818 Patentiasan 2 NL — 2230 HV Rijsevijk Tel. (-31-70) 340-2040, Tx. 31 651 spo rd, FISE: (-331-70) 340-3016	Authorized officer Mand I , B	

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INTERNATIONAL SEARCH REPORT

Int Sonal Application No PCT/US 99/25694

A Continue	INTO DOCUMENTS CONSIDERED TO BE RELEVANT	PCT/US 99/25694
	Citation of document, with indication,where appropriate, of the relevant passages	- In
,	Annual Advisor of the second o	Relevant to claim No.
P,X	XIAO W. ET AL.: "Gene therapy vectors based on adeno-associated virus type 1." JOURNAL OF VIROLOGY, vol. 73, no. 5, May 1999 (1999-05), pages 3994-4003, XP002137090 ISSN: 0022-538X the whole document	1-23
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INTERNATIONAL SEARCH REPORT

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1. X Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely: Remark: Although claims 18-20 and 22, as far as an in vivo application is concerned, are directed to a method of treatment of the human or animal body, the search has been carried out and based on the alleged effects of the compound/composition.
Claims Nos.: because they relate to parts of the international Application that do not comply with the prescribed requirements to such an extent that no meeningful international Search can be carried out, specifically:
Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box ii Observations where unity of invention is lacking (Continuation of Item 2 of first sheet)
This international Searching Authority found multiple inventions in this international application, as follows:
As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
As only some of the required additional search fees were timely paid by the applicant, this international Search Report covers only those claims for which fees were paid, specifically claims Nos.:
No required additional search fees were timely paid by the applicant. Consequently, this international Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remark on Protest The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees.

. INTERNATIONAL SEARCH REPORT

information on patent tendly members

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Patent document cited in search report		Publication date	P	etent family nember(s)		Publication date		
WO 9811244	A	19-03-1998	AU EP	46456 09326	97 A 94 A	02-04-1998 04-08-1999		
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